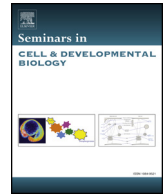




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## Review

## Neural stem cell niche heterogeneity

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## HIGHLIGHTS

- Neural stem cells are not equally plastic homogeneous cells, but rather a combination of distinct subpopulations.
- The cerebrospinal fluid is an essential component of the neural stem cell niche.
- Neural stem cells auto-regulate themselves.
- Innervations release neurotransmitters to neural stem cells, and affect neural stem cell behavior.
- The biggest challenge remains to study the neural stem cell niche in humans.

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## ABSTRACT

In mammals, new neurons can be generated from neural stem cells in specific regions of the adult brain. Neural stem cells are characterized by their abilities to differentiate into all neural lineages and to self-renew. The specific microenvironments regulating neural stem cells, commonly referred to as neurogenic niches, comprise multiple cell populations whose precise contributions are under active current exploration. Understanding the cross-talk between neural stem cells and their niche components is essential for the development of therapies against neurological disorders in which neural stem cells function is altered. In this review, we describe and discuss recent studies that identified novel components in the neural stem cell niche. These discoveries bring new concepts to the field. Here, we evaluate these recent advances that change our understanding of the neural stem cell niche heterogeneity and its influence on neural stem cell function.

**Abbreviations:** 3V, third ventricle; ARC, arcuate nucleus; CD133, cluster of differentiation 133, also known as prominin-1; CD15, cluster of differentiation 15, also known as 3-fucosyl-N-acetyl-lactosamine; CD24, cluster of differentiation 24, also known as Heat Stable Antigen; CD81, cluster of differentiation 81, also known as target of the antiproliferative antibody 1 and tetraspanin-28; CD9, cluster of differentiation 9, also known as the motility related protein-1; Cre, recombinase; CreER, recombinase binded to estrogen receptor; CSF, cerebrospinal fluid; DCX, doublecortin; DG, dentate gyrus; DMH, dorsomedial hypothalamic; DREADDs, Designer Receptors Exclusively Activated by Designer Drugs; EGFR, Epidermal growth factor receptor; ERT2, mutated ligand-binding domain of the estrogen receptor; FGF-2, fibroblast growth factor type 2; GABA, gamma-aminobutyric acid; GABAergic, pertaining to or affecting the neurotransmitter GABA; GCL, granule cell layer; GFAP, glial fibrillary acidic protein; Glut, glutamate aspartate transporter; Gli1, GLI-, Kruppel family member 1, also known as glioma-associated oncogene; IGF-1, Insulin-Like Growth Factor 1; L-DOPA, L-3,4-dihydroxyphenylalanine, also known as levodopa; LHA, lateral hypothalamic area; LoxP, locus of X-over P1; ME, median eminence; Mfge8, milk fat globule-epidermal growth factor, also known as lactadherin or SED1; mRNA, messenger ribonucleic acid; mTOR1, mammalian target of rapamycin 1; NeuN, neuronal nuclear antigen; NG2, neuron-glia antigen 2, also known as CSPG4; NSC, neural stem cell; PSA-NCAM, Polysialylated neural cell adhesion molecule; PVH, paraventricular nucleus; RGLs, radial glia like neural stem cells; RNA, ribonucleic acid; RNAseq, ribonucleic acid sequencing; SGZ, subgranular zone; SGZ, subgranular zone; Sox1, SRY-related HMG-box gene 1; SVZ, subventricular zone; Tie2, tyrosine kinase with immunoglobulin-like and EGF-like domains 2; VE-Cadherin, vascular endothelial cadherin; VEGF, vascular endothelial growth factor; VIP, vasoactive intestinal peptide; VMH, ventromedial hypothalamic; Wnt, wingless/integrated;  $\alpha$ -SYN,  $\alpha$ -synuclein

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## 1. Introduction

Since the time of Santiago Ramón y Cajal, for almost a century, the adult nervous tissue had been mistakenly considered invariable in all animals. This concept started changing, when, initially, it was discovered that invertebrates are not included in this conception, especially during metamorphosis [1]. Later, also lower vertebrates, such as birds during their song learning process, were described to possess plasticity in the central nervous system [2]. Finally, in mammals, while the vast majority of neurons are generated in utero [3], at the end of last century, neural stem cells were described in specific regions in the adult brain [4].

Neural stem cells are undifferentiated neural cells that are defined based on their extensive replicative potential, their ability to differentiate into multiple central nervous system neuronal and glial cell types, and their capacity for long-term self-renewal. Albeit the high proliferative capacity is a hallmark of stemness, a unique characteristic of neural stem cells, comparing to other central nervous system cells, is their capability to stay dormant for very long periods, providing a reserve pool of cells available for tissue regeneration and cell replacement throughout life [5,6].

In the adult mammalian brain, there are at least two areas that are neurogenic and contain a reservoir of neural stem cells: the subgranular zone in the hippocampal dentate gyrus and the subventricular zone around the lateral ventricles [7]. Interestingly, the hypothalamus has been recently identified as a possible third neurogenic area in the mammalian brain [8,9] (Fig. 1). Hypothalamic neurogenesis has been associated to the regulation of body weight homeostasis and the control of energy balance [10,11]. Quiescent and activated neural stem cells coexist within these neurogenic regions generating new cells throughout life [12–14]. The role of adult neurogenesis goes also

beyond the simple replacement of cell loss in the adult central nervous system, as it has been associated with multiple brain functions.

Adult neural stem cells in the dentate gyrus originate intermediate progenitor cells [15], which go through fast, but limited, divisions before they exit the cell cycle and differentiate into mature astrocytes and neurons. This process has been demonstrated to be crucial in memory formation and behavioral performance [16]. Interestingly, physical exercise, which promotes learning and memory, activates subgranular zone neurogenesis [17–19]. Within the adult subventricular zone, activated neural stem cells form neuroblasts which travel through the rostral migratory stream to the olfactory bulb, where they originate periglomerular and granule mature neurons [20]. These neurons formed in the olfactory bulb are involved in olfactory learning during adulthood [21,22]. Adult neural stem cells also originate NG2-glia cells that disperse to the white and gray matter, which can generate corpus callosum oligodendrocytes or olfactory bulb interneurons [23–25].

It is still unclear how neural stem cells are actively maintained throughout life, and what are the cellular interactions, molecular cascades, and accountable cell and non-cell-autonomous signals that regulate neural stem cell behavior. Tissue microenvironments are very complex, being composed of multiple cell types with a variety of cues that are constantly released [26–32,186,187]. Understanding the signaling mechanisms that determine neural stem cell fate will be crucial for the success of clinical applications targeting these cells. Recent studies using genetically modified mouse models indicate that the fate of neural stem cells is finely regulated by changes in the surrounding microenvironment, also termed *niche*, in which they reside. These changes are dictated by both extrinsic (for instance, physical activity, stress, environmental enrichment, or aging) and intrinsic (for instance, cytokines, growth factors, hormones, or neurotrophins) factors [33]. Therefore, decisions regarding neural stem cell self-renewal, activation

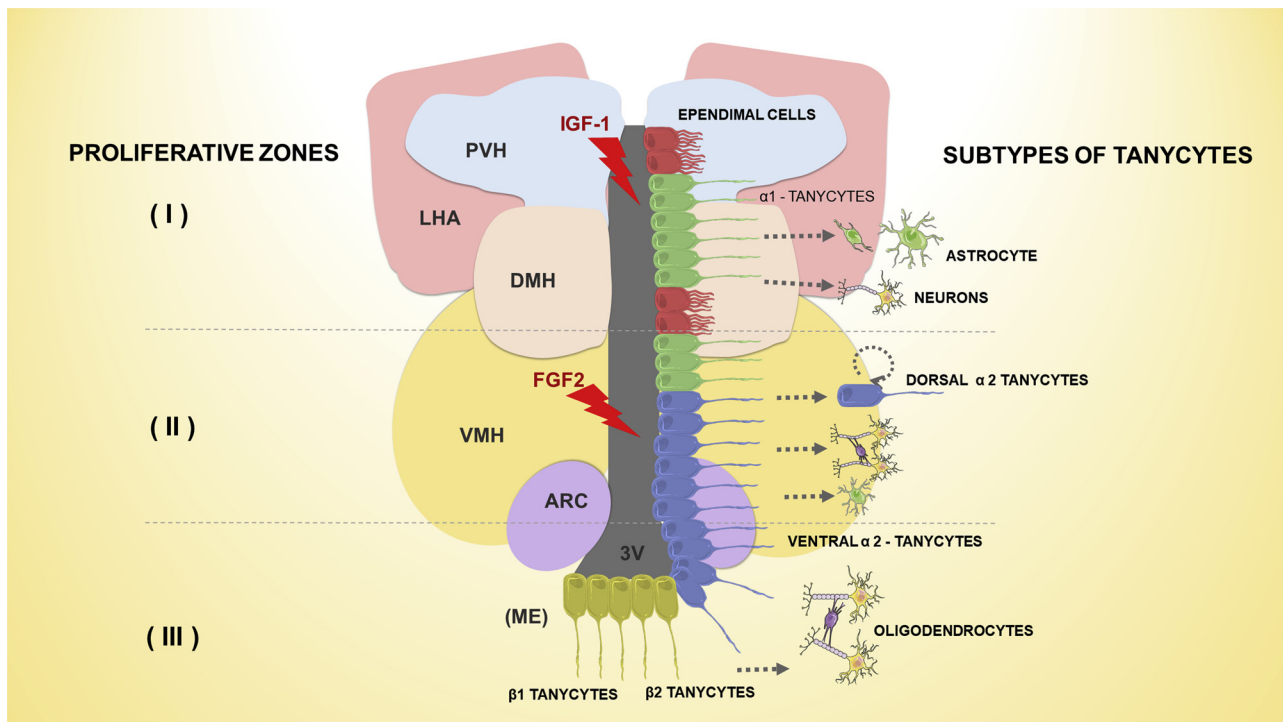


Fig. 1. Presence of neural stem cells (tanyocytes) in the adult hypothalamus.

Three proliferative zones were reported in the adult hypothalamus. The first (I) proliferative zone, in the dorsal  $\alpha 1$  region, after a stimulus (IGF-1) can induce tanyocytes to originate neurons and possibly astrocytes in the adjacent hypothalamic parenchyma. The second (II) proliferative zone, in the dorsal  $\alpha 2$  region, can be stimulated by FGF-2 activating the symmetric self-renewal of dorsal  $\alpha 2$  tanyocytes or can give origin to neurons and astrocytes; and rarely oligodendrocytes. In the third (III) proliferative zone, called “hypothalamic proliferative region”, located in the adjacent median eminence (ME), tanyocytes proliferate symmetrically giving rise to neurons, and possibly progenitors of oligodendrocytes. FGF-2: fibroblast growth factor type 2; IGF-1: Insulin-Like Growth Factor 1; PVH: paraventricular nucleus; LHA: lateral hypothalamic area; DMH: dorsomedial hypothalamic; VMH: ventromedial hypothalamic; ARC: arcuate nucleus; 3V: third ventricle.

or differentiation are dependent on the interaction with constituents from their niche. The deregulation of those microenvironmental regulatory mechanisms may cause dysfunction of neural stem cells, leading to neurological disorders [34].

The discovery of neural stem cells in the adult brain provided us with a promising target for central nervous system disease therapies [35]. A large number of investigations were already performed in order to understand the behavior of these cells in the adult brain; nevertheless, the best is yet to come. Recently, several components of the neural stem cell niche have been identified, regulating neural stem cell activity by supplying various signals. In this review, we present an overview of the current knowledge on the variety of brain components in the neurogenic niche and their effects on neural stem cells.

## 2. Heterogeneity of neural stem cells

Neural stem cells are not equally plastic homogeneous cells, but rather a combination of distinct subpopulations [36]. This concept needs to be considered to fully understand the relationship between adult neural stem cells and their niches. Neural stem cells display regional heterogeneity possibly acquired from their embryonic origin and niche patterning [188]. Viral lineage-tracking and recombinase-based fate mapping experiments of cell populations in distinct dorso-ventral or rostral-caudal regions of the adult subventricular zone revealed that a mosaic of neural stem cells are distributed in diverse domains, correlating with specific regional expression of particular transcription factors [37]. This subventricular zone regional identity of adult neural stem cells appears as early as embryonic day E15.5 [38]. Nevertheless, it remains poorly explored how exactly adult neural stem cells become regionally specified. Interestingly, heterotopic transplantation studies suggest that neural stem cell identity is partially a cell intrinsic characteristic, as neural stem cells, after transplant to a different neurogenic area, keep their regional identity and continue to produce the same progeny as in the original position [39]. Additionally, neural stem cells undergo changes in chromatin structure, mRNA, and noncoding RNA levels that make them more or less sensitive to external signals over short time periods [40–42].

The unique genetic signature reflects the regional identity of neural stem cells. Importantly, the heterogeneity revealed at the molecular level may translate into singular functional differences. Both quiescent and activated neural stem cells are present within the neurogenic niches [5]. Multiple molecular markers were proposed to be used to distinguish neural stem cell subsets such as CD15 [43], CD133 [44], Sox1 [45], Nestin [46–48], and EGFR [49]. More recently, the possibility of analyzing multiple molecular markers in combination (GFAP, EGFR, CD133, Nestin, CD9, CD81, CD24, and VEGF), by the use of transgenic mice, flow cytometry, and single cell RNAseq, revealed the complexity within the neural stem cells population [50–53]. These differences within neural stem cells possibly reflect transcriptional networks and signaling set points unique to subsets of neural stem cells. Although the regional identity of neural stem cells has been mainly typified in mouse models, analyses in the primate brain have also revealed heterogeneity of subventricular zone neural stem cells which declines with aging [54]. Nevertheless, our knowledge on the human neural stem cells heterogeneity remains very limited. In the future, deciphering the functional consequences of adult neural stem cell heterogeneity will be crucial to understand brain functioning in physiologic and pathologic conditions.

Although neural stem cell heterogeneity has not been exhaustively explored yet in the adult dentate gyrus, subpopulations of neural stem cells with different morphologies and behaviors have been described in this neurogenic area [55,56]. The dentate gyrus is anatomically subdivided into temporal and septal regions [57]. Remarkably, these two areas differ in their functionality and molecular composition [58]. There is a gradient in the expression of several molecules throughout the dentate gyrus. For instance, Wnt inhibitor Frizzled-related protein 3 is highly expressed in the temporal region in comparison with the

expression in the septal area [59]. Interestingly, genetic deletion of this gene results in activation of neural stem cells in the temporal region, indicating a potential molecule that creates spatial heterogeneity in the adult dentate gyrus [60]. Moreover, neurogenesis in the septal region, in comparison with the temporal region, is faster [61]; while the density of neural stem cells in the temporal region is lower than in the septal region [62]. Unfortunately, still there are no defined molecular markers to distinguish neural stem cell subtypes *in vivo*.

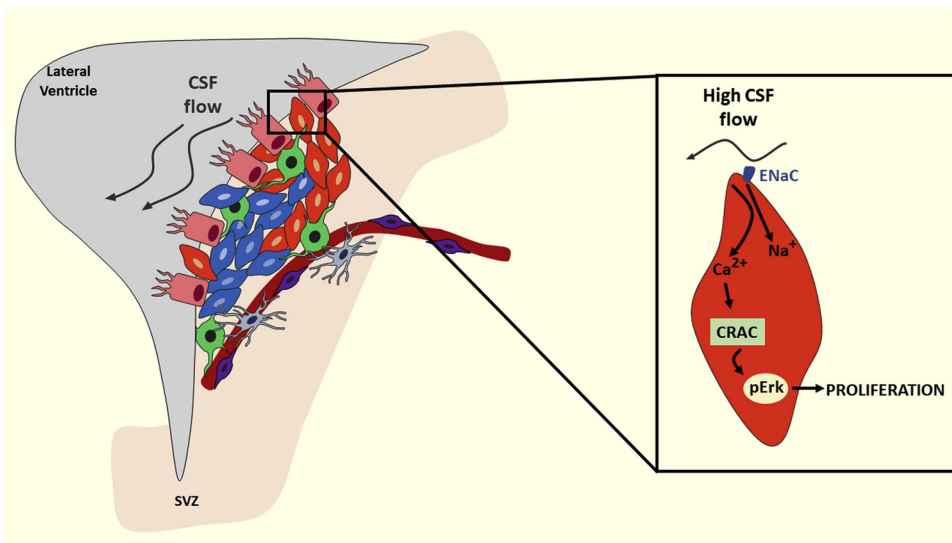
In a recent elegant study, Song's group used morphological differences to separate subgranular zone neural stem cells into two subsets [56]. The rarer neural stem cells, termed type  $\beta$  cells, have shorter and more branched processes compared to the more abundant type  $\alpha$  cells. Genetic fate mapping revealed that type  $\alpha$  cells can originate type  $\beta$  cells, therefore being hierarchically superior to type  $\beta$  cells, while type  $\beta$  cells did not proliferate. Nevertheless, the heterogeneity within the population of type  $\alpha$  cells was not yet explored, and remains unknown whether all type  $\alpha$  cells can form type  $\beta$  cells. These data suggest that there are different neural stem cell populations in the adult dentate gyrus. Single cell RNAseq analyses will permit high throughput data collection that may reveal whether these cell populations can be further subdivided into multiple subsets with distinct functions, reacting differently to niche signaling.

Novel tools should be generated to determine what constitutes a population of neural stem cells versus transition cellular states. As neural stem cells also present microenvironment dependent plasticity, future studies should explore how neural stem cells heterogeneity affects cell competition within the same niche, and whether such heterogeneity allows adaptation to distinct microenvironmental cues. Defining the signals that can influence different neural stem cells populations behavior will have important implications for developing therapeutic strategies for neural disorders based on the mobilization of endogenous neural stem cells.

Very little is known about the extrinsic signals that regulate neural stem cells subpopulations. It is still not completely known whether and how the neurogenic microenvironments differ. The functional heterogeneity of neural stem cells points to the potential for matching heterogeneity of influences from the niche that support the behavior of these neural stem cell subsets. The remainder of this review will focus on the neural stem cell niche heterogeneous components.

## 3. Cerebrospinal fluid in the neural stem cell niche

Neural stem cells in the subventricular zone are in direct contact with the cerebrospinal fluid-filled ventricles [63]. The cerebrospinal fluid is a reservoir of signaling molecules, which is modulated by external factors. Subsequently, neural stem cells receive cues from this fluid, allowing them to sense external changes. The cerebrospinal fluid is essential for the development and maintenance of the central nervous system [64–66]. It contains multiple trophic factors essential for the maintenance and proliferation of neural stem cells, including trophic factors and neuroendocrine peptides [67–75]. Cerebrospinal fluid also regulates neural stem cell behavior via hydrostatic forces [76]. Now, in a recent article in *Cell Stem Cell*, Petrik and colleagues reveal a molecular sensor present in neural stem cells which detects cerebrospinal fluid flow [77] (Fig. 2). The authors demonstrated that neural stem cells are highly enriched with the epithelium sodium channel [77]. Petrik and colleagues investigated the role of the epithelium sodium channel in neural stem cells by using state-of-the-art techniques, including subventricular zone whole mounts, sophisticated Cre/loxP techniques *in vivo*, confocal microscopy, and electrophysiological recordings. These experiments showed that epithelium sodium channel is essential for neural stem cells proliferation *in vitro* [77]. Strikingly, genetic deletion of epithelium sodium channel specifically from neural stem cells reduced the number of neural stem cells and inhibited their activation and proliferation, leading to reduction of neuroblasts derived from these neural stem cells [77]. Interestingly, artificial cerebrospinal fluid



**Fig. 2.** Changes in cerebrospinal fluid flow can alter neural stem cell proliferation and neurogenesis. In the subventricular zone (SVZ) of the lateral ventricles, neural stem cells are in close contact with the cerebrospinal fluid (CSF). Alterations in the cerebrospinal fluid flow are detected by epithelial sodium channels present in these cells, which can affect their proliferation and differentiation. Epithelial sodium channel (ENaC), calcium release-activated channels (CRAC), phosphorylated extracellular signal-regulated kinases (pErk).

flow promoted neural stem cells proliferation in the whole mount subventricular zone through epithelium sodium channels. As when these channels were specifically blocked, the effect on neural stem cells proliferation disappeared [77]. Additionally, Petrik and colleagues demonstrated that within neural stem cells epithelium sodium channels induce sodium and calcium influx in response to cerebrospinal fluid flow [77].

This study reveals a novel mechanism of the interaction between cerebrospinal fluid flow and neural stem cells, nonetheless several questions remain unanswered. Conditional gene manipulation strategies, as the ones used in this study, offer a powerful tool to study the role of specific genes in particular cell populations [78]. Nevertheless, this type of studies also may have their caveats. The main findings from this study are based on the data obtained from tamoxifen-inducible *Glast* promoter driven *CreER-LoxP* system. In this mouse model, cells produce the recombinase based on their expression of the glutamate aspartate transporter (*Glast*). Albeit neural stem cells express *Glast*, they are not the only cells in the brain that express this gene, and, more importantly, other cells present in the subventricular zones will produce the recombinase in those mice as well, such as pericytes [79]. Therefore, it remains to be answered whether the genetic ablation of epithelium sodium channel in *Glast*-expressing cells may contribute to the behavior of neural stem cells indirectly in the subventricular zone. Another question that remains open, is, as discussed above, that neural stem cells are heterogeneous in their niche. Future studies should examine whether epithelium sodium channel is crucial to sense the cerebrospinal fluid flow in all neural stem cells or in a specific subpopulation. It remains also unknown whether the importance of this channel in sensing the cerebrospinal fluid flow is restricted to a certain time period throughout life, and when during development this channel becomes crucial.

Importantly, it still needs to be explored whether the role of the epithelium sodium channel on neural stem cells is altered in neurodegenerative disorders. Are some of the defects in neurogenesis seen in brain pathologies due to altered sensing of the cerebrospinal fluid flow by neural stem cells? As neural stem cells are key in neural regeneration, is the neurogenic control by the cerebrospinal fluid flow transmitting physiological and pathological conditions of the brain to the neural stem cells?

#### 4. Autocrine regulation in the neural stem cell niche

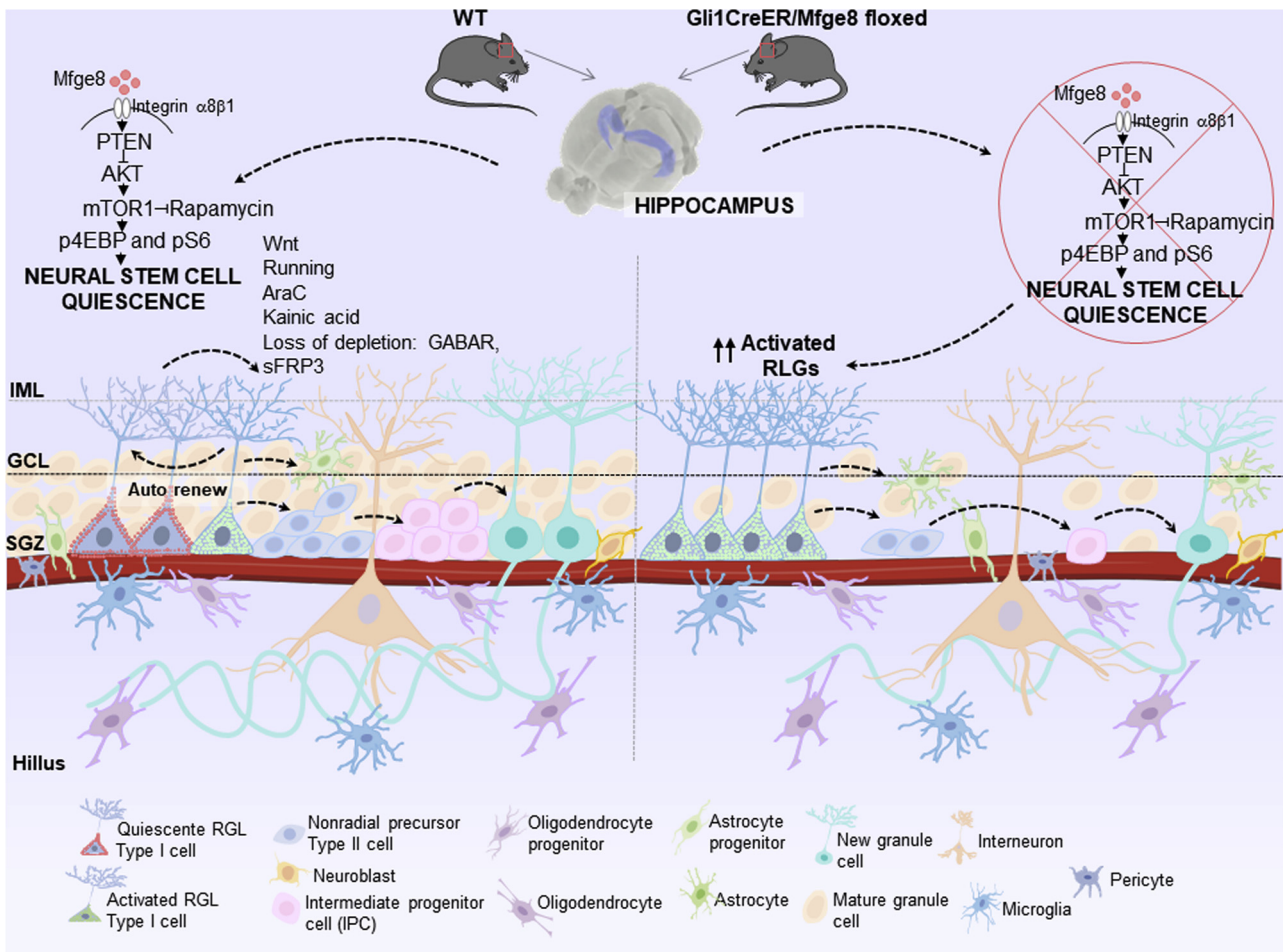
Multiple signaling molecules identified in the neurogenic niche regulate the behavior of neural stem cells, such as neurotransmitters,

growth factors, and membrane-associated ligands. Most of these molecules are derived from various cellular components of the neural stem cell niche, and act in a paracrine, non-cell autonomous, way. Whether neural stem cells can be regulated by factors derived from themselves remains unknown. Now, in a recent article in *Cell Stem Cell*, Zhou and colleagues show that neural stem cells in the subgranular zone can regulate their own quiescence in an autocrine manner [80] (Fig. 3). Interestingly, based on a single neural stem cell RNAseq analysis from the same group [81], the authors found that lactadherin, also known as *Mfge8* or *SED1*, is highly expressed in quiescent neural stem cells from the dentate gyrus. Zhou and colleagues used state-of-the-art techniques, including sophisticated *Cre/loxP* technologies, *in vivo* lineage-tracing, and confocal microscopy to determine the role of lactadherin in neural stem cells in the subgranular zone. Their results demonstrated that lactadherin is required to promote the maintenance and quiescence of neural stem cells in the hippocampus. Strikingly, genetic ablation of lactadherin from neural stem cells led to decreased density of quiescent neural stem cells in the adult subgranular zone [81]. Furthermore, short-term fate tracking upon deletion of lactadherin in quiescent neural stem cells revealed a significant increase in proliferating neural stem cells, indicating that lactadherin blocks neural stem cell activation and proliferation. Finally, Zhou and colleagues explored the mechanism by which lactadherin promotes neural stem cell quiescence, demonstrating that it is via suppression of *mTOR1* pathway [80]. This work provides a novel role for lactadherin in the hippocampus, and reveals that neural stem cells are also crucial in the formation of their own niche.

Zhou and colleagues examined neural stem cell as a homogeneous cell population in their study [80]. Nevertheless, as mentioned above, it would be interesting to explore whether lactadherin is restricted to a neural stem cell subset. Moreover, the mouse model that was used to study neural stem cells (*Gli1-CreERT2* mice) is not specific to neural stem cells, as it presents recombinase activity also in perivascular cells [82–84]. Therefore, future experiments will reveal whether lactadherin is an exclusively autocrine signal or if it may derive from other niche components as well. Additionally, it remains unknown whether in other neurogenic niches besides the hippocampus lactadherin has a similar role.

#### 5. Perivascular neural stem cell niche

Stem cells from multiple organs are located in a very close position to the vascular network, including neural stem cells, implicating that blood vessels are an integral constituent of the stem cell niche



**Fig. 3.** Autocrine signaling in the subgranular zone (SGZ) in the dentate gyrus. Quiescent radial glia like neural stem cells (RLGs) are activated and continuously give rise to newborn dentate granule cells. Milk fat globule-epidermal growth factor (Mfge8), also called lactadherin, is a neural stem cell enriched niche factor that maintains the neural stem cell pool in the dentate gyrus during early postnatal development and in the adulthood by promoting neural stem cell quiescence. Mfge8 is enriched in quiescent neural stem cell and regulates neural stem cell quiescence, via mTOR1 signaling, and its deletion depletes neural stem cells and decreases adult neurogenesis.

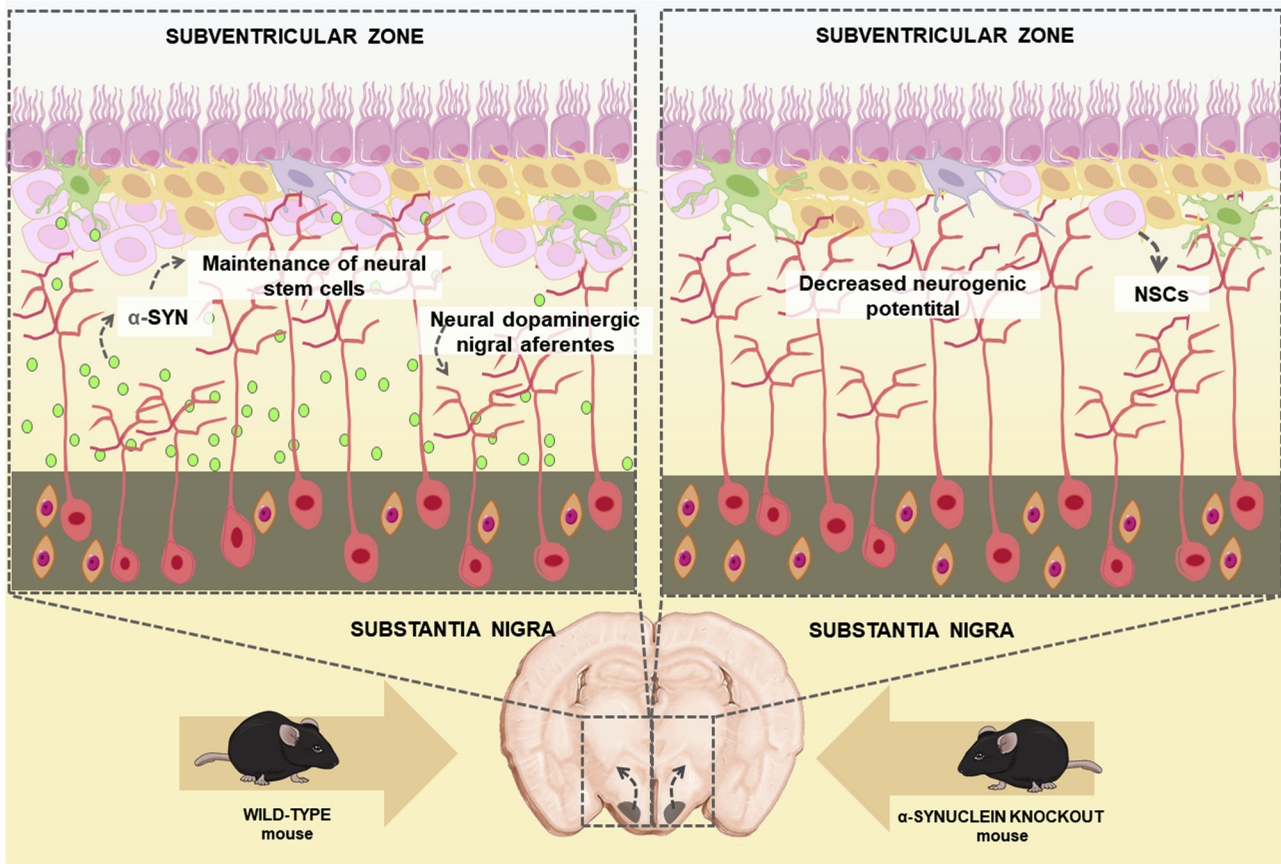
[24,85–91]. Interestingly, the neurogenic regions are more vascularized than not-neurogenic areas in the brain [91–93]. The vasculature in these regions is more permeable, and the blood flow is slower, suggesting that there is facilitation in the access of blood-derived signals to the neural stem cells [24,91,92,94]. In both main neurogenic regions, subventricular and subgranular zones, proliferating neural stem cells are in close physical proximity to the vasculature than other niche cellular components [89,86–91,95,96]. Importantly, transplanted neural stem cells also preferentially associate to blood vessels [97]. Although some studies correlate angiogenesis in the neural stem cell niche with increased neurogenesis [98–101,18,102,103], others show that expansion of the niche vasculature not necessarily is accompanied by augment in neurogenesis [98].

Demonstrating the functional significance of endothelial cells in the neural stem cell niche, Otonne and colleagues have demonstrated functional contact mediated cross-talk between endothelial cells and neural stem cells *in vivo* [104]. Conditional deletion of the transmembrane ligands, Jagged1 or ephrinB2, specifically in endothelial cells *in vivo* culminates in an expansion of proliferating neural stem cells, followed by their long-term depletion [104], indicating that these molecules are crucial to keep a reserve of neural stem cells in a quiescent state.

In the search for endothelially-secreted factors that regulate neural stem cells behavior, a recent study by Sato et al. (2017) revealed that soluble amyloid precursor protein derived from endothelial cells is

essential for neural stem cell quiescence [105,106]. The authors found by *in vitro* experiments that soluble amyloid precursor protein suppresses neural stem cell growth, and enhances neurosphere-forming capacity, while maintaining their multipotency. Moreover, Sato and colleagues also discovered that, in amyloid precursor protein-null mice, there is a rise in neural stem cells proliferation in their niche [105]. Furthermore, using Tie2-Cre/amyloid precursor protein floxed mice, the authors deleted amyloid precursor protein specifically in endothelial cells, revealing that endothelial cells, but not astrocytes regulate neural stem cells activation in the subventricular zone through amyloid precursor protein [105]. As Tie2 expression is not exclusive to endothelial cells [107,108], it is possible that the observed neural stem cell response may be due to hematopoietic cells in which amyloid precursor protein was also genetically eliminated as well in Tie2-Cre/amyloid precursor protein floxed mice. To avoid Cre recombinase activity in hematopoietic cells, more specific mouse models should be used in future studies, such as VE-Cadherin-CreERT2 mice [109]. In VE-Cadherin-CreERT2/amyloid precursor protein floxed is possible control amyloid precursor protein expression in the endothelium at different stages.

The perivascular niches are themselves complex and heterogeneous composed by multiple other cell types in addition to endothelial cells in the neurogenic niches, such as perivascular astrocytes, perivascular neurons [106,110,111], perivascular macrophages [112–114], perivascular adventitial cells [115], perivascular fibroblasts [116],



**Fig. 4.**  $\alpha$ -synuclein ( $\alpha$ -SYN) present in dopaminergic nigral afferents is essential for the normal cycling and maintenance of neural stem cells (NSCs) in the adult subventricular zone.

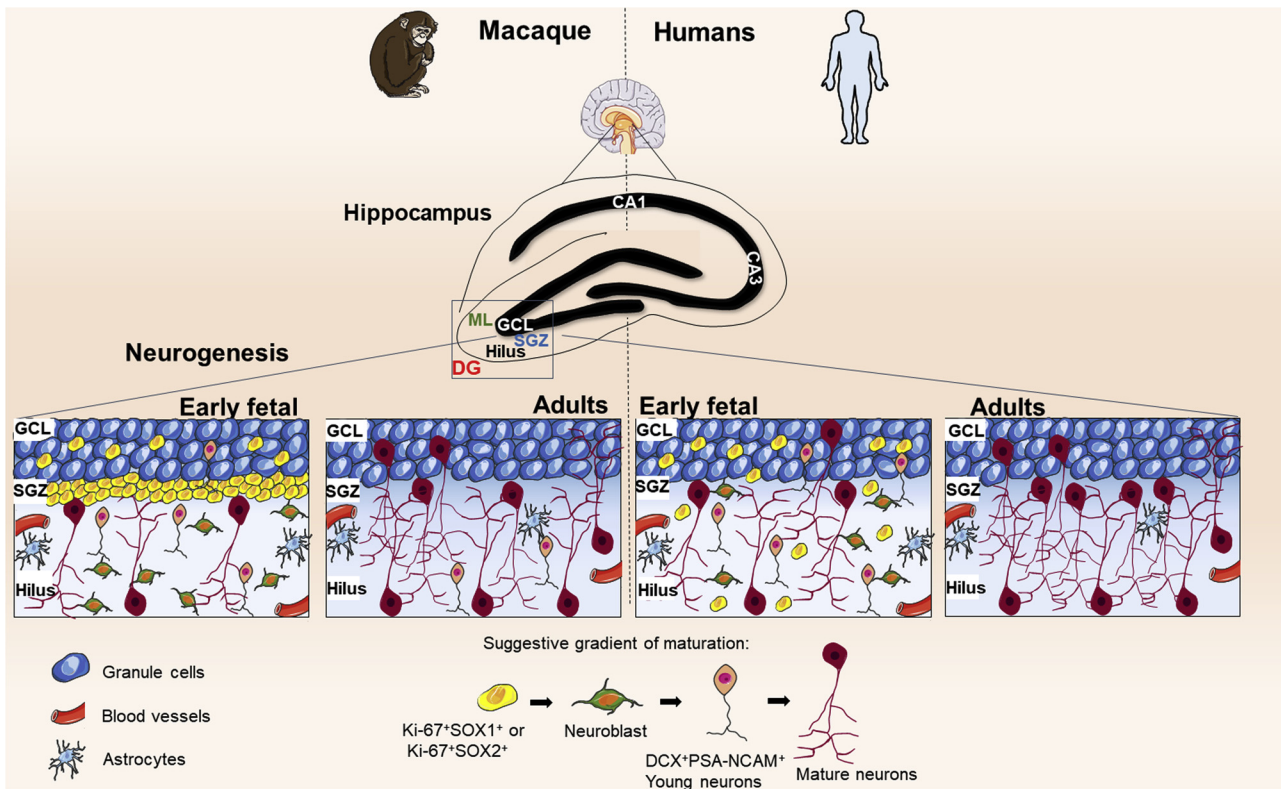
perivascular microglia [117], vascular smooth muscle cells [118], and pericytes subsets [119]. Interestingly, pericytes from several peripheral tissues have been shown to behave as stem cells [78,120–139,189,190], generating other cell types, and also to regulate the behavior of other stem cells, as hematopoietic stem cells in their niches [87,88,140–144]. Although growing evidence also shows that central nervous system pericytes alter their characteristics following stimuli and develop stemness [145–149], whether pericytes are an essential component of neural stem cell niche, and what are their exact roles remains unexplored. Future studies should elucidate what is the relationship between different components of the perivascular neural stem cell microenvironment. Is there a cross-talk between the different perivascular constituents of the neural stem cell niche? Also, the identification of molecules contributing to the anchoring of neural stem cells to the perivascular niche deserves further research.

## 6. Neuronal role in the neural stem cell niche

Neurotransmission has been classically defined as signaling between neuronal subtypes. Nevertheless, this concept has changed, and now we know that released neurotransmitters can signal not only to neurons, but also to other cell types present in the microenvironment where they are released. Recent studies have revealed that innervations are essential components of the neural stem cell niche, and the release of neurotransmitters affects the neural stem cell behavior in the neurogenic area [150]. Different from embryonic neurogenesis, adult neurogenesis is regulated also by neuronal activity [3,33,151–154]. Understanding which are the specific neurons that regulate neural stem cells in the adult brain and the underlying mechanisms is fundamental yet challenging. Mostly due to the lack until recently of techniques to access the behavior of neural stem cells in vivo. The recent technical

advancements, such as characterization of specific neural stem cell markers, DREADDs (designer receptors exclusively activated by designer drugs), optogenetics, cross-synaptic specific tracing, and the evolution of Cre/lox systems are bringing exciting findings and novel concepts to the field. Now researchers are in a unique position to answer essential questions about neuronal activity-dependent neural stem cell regulation.

Using the combination of lineage-tracing and optogenetics, Song and colleagues have recently revealed that dentate parvalbumin + interneurons regulate adult hippocampal neural stem cells via tonic GABA signaling in an activity-dependent manner [155,156]. Decrease in the activity of these interneurons promotes increase in the number of quiescent neural stem cells, while conversely activation of parvalbumin + interneurons inhibits the activation of neural stem cells [152,155]. Interneurons expressing neuropeptide Y have been implicated in the control of adult neurogenesis via promoting neural stem cell proliferation [157–159]. Moreover, interneurons expressing vasoactive intestinal peptide (VIP) have been shown to mediate neurogenesis via VIP receptors on neural stem cells [160]. As hippocampal interneurons receive afferent inputs from distant brain regions, there is a need to understand how distal neuronal inputs impact adult hippocampal neural stem cells via local interneurons. One recent study by Bao and colleagues (2017) addressed this by using state-of-art techniques [161]. The authors discovered by using virus-based retrograde tracing that medial septum GABAergic neurons are the main afferents to the hippocampal parvalbumin + interneurons [161]. Markedly, GABA signaling derived from medial septum GABAergic neurons onto hippocampal parvalbumin + interneurons leads to neural stem cell regulation [161]. Depletion of medial septum GABAergic neurons results in neural stem cell depletion in the hippocampus, indicating that distal brain activity regulates hippocampal neural stem cell behavior



**Fig. 5.** Hippocampal neurogenesis was not detected in adult humans by Sorrells et al. (2018). The study of Sorrells and colleagues suggests differences in neurogenesis among primates. In humans, in the dentate gyrus (DG), a proliferative subgranular zone (SGZ) is not formed near the granule cell layer (GCL), instead, the proliferating cells, which express progenitor stem markers, are mostly scattered in the hilus and depleted from the 7 years old. The number of young neurons, DCX + PSA-NCAM + cells, in GCL and hilus, also decrease from birth and in adult individuals, these cells are no longer found in the hippocampus. In adult humans, the predominance of morphologically mature neurons expressing PSA-NCAM + and NeuN was observed. In rhesus macaque (*M. mulatta*) there are some differences in the process of neurogenesis. In DG the formation of a germinative proliferative subgranular zone (SGZ) is observed. The number of proliferative cells and young neurons DCX + PSA-NCAM + decreases until the 7 years old, an age in which the germ cell layer in the SGZ already becomes dispersed. A developmental normalization has shown that the decrease of young neurons in humans is more accelerated than in monkeys, which allows the identification of still rare DCX + PSA-NCAM + neurons in adult monkeys. PSA-NCAM: Polysialylated neural cell adhesion molecule; DCX: doublecortin; NeuN: neuronal nuclear antigen.

[161]. Detailed characterization of multiple synaptic inputs that end onto distinct cellular components of the hippocampal neural stem cell niche will provide insights into the spatial organization of the local circuitry, and how it affects neural stem cell niche constituents.

The subventricular zone is also innervated by multiple nerve fibers of different origins that may influence the neural stem cell behavior. Neurons expressing nitric oxide synthase regulate neural stem cell proliferation by nitric oxide production [162]. The subventricular zone is also supplied by choline acetyltransferase expressing axons which regulate neural stem cell proliferation through the neurotransmitters that they produce [163,164]. Additionally, selective lesion of dopaminergic nerve fibers leads to reduced proliferation of neural stem cells in the subventricular zone [165–168]. In a recent article in *Journal of Neuroscience*, Perez-Villalba and colleagues revealed an important component of the subventricular zone neural stem cell niche:  $\alpha$ -synuclein possibly derived from dopaminergic axons maintains neural stem cells in their subventricular niche [169] (Fig. 4). The authors identified that  $\alpha$ -synuclein is expressed in dopaminergic nerve fibers innervating the subventricular zone, but is not present in the subventricular zone itself. Perez-Villalba and colleagues discovered that the absence of  $\alpha$ -synuclein, in  $\alpha$ -synuclein knockout mice, leads to reduction in neural stem cells in the subventricular zone. Strikingly, adenovirus-mediated expression of  $\alpha$ -synuclein in the substantia nigra neurons and L-DOPA treatment prevent neural stem cell loss in the neurogenic area [169]. This study brings strong evidence for the participation of dopaminergic in the maintenance of adult neural stem cells. Nevertheless, further studies need to confirm the participation of  $\alpha$ -synuclein derived from

dopaminergic fibers in the neural stem cell niche. The use of conventional knockout mice has proved to be a valuable tool for understanding the role of key proteins in physiological and pathological states. Nonetheless, these technologies produce broad changes in gene function throughout the body, affecting multiple different cells. Thus, they are limited in that they do little to identify the specific roles of a gene in a specific cell type. Because the molecular functions of  $\alpha$ -synuclein may depend on a specific neuronal subpopulation in which it is expressed, restricting gene manipulation to specific neurons in the brain may be more useful for understanding the role of  $\alpha$ -synuclein in the neural stem cell niche. Thus, conditional gene manipulation approaches provide an effective option. The main findings from this study are based on the data obtained from  $\alpha$ -synuclein knockout mice. As during development  $\alpha$ -synuclein may be expressed in distinct tissues in various cellular populations [170], it is possible that the effect on neural stem cells could be due to other cell types in which  $\alpha$ -synuclein was deleted as well in the  $\alpha$ -synuclein knockout mouse model. Because of this, the combination of mouse models that allow to target specifically dopaminergic neurons, such as tyrosine hydroxylase-Cre, with  $\alpha$ -synuclein floxed mice will provide a tool to study the role of  $\alpha$ -synuclein specifically within dopaminergic neurons.

Interestingly, recently, in *Science*, Paul and colleagues demonstrated that a subpopulation of hypothalamic nerve fibers essential in the control of hunger and satiety regulate adult neural stem cells proliferation *in vivo* [171,172]. The authors identified by *in vitro* assays  $\beta$ -endorphin, an endogenous kappa opioid receptor ligand, as an extrinsic cue that activate quiescent neural stem cells. Furthermore, as  $\beta$ -

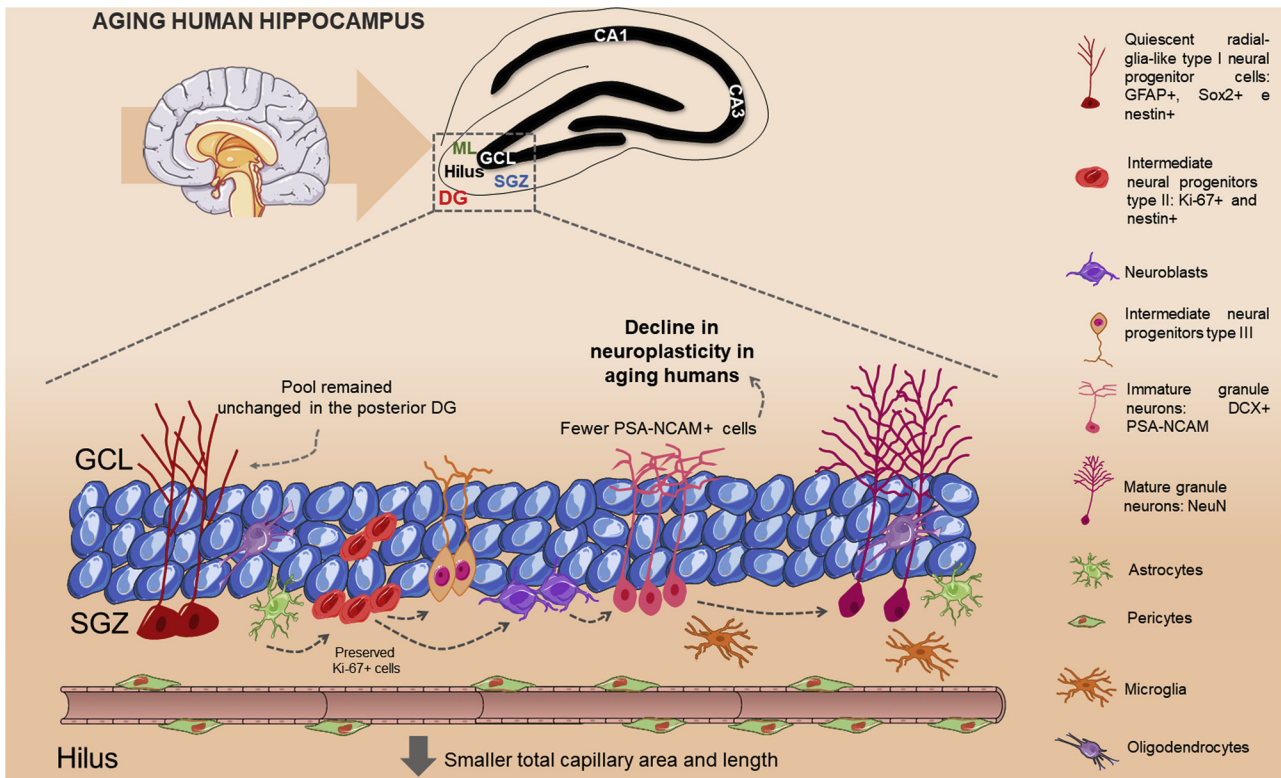


Fig. 6. Hippocampal neurogenesis continues during aging in humans.

The study of Boldrini and colleagues suggests that neurogenesis is present in the human hippocampus. PSA-NCAM was detected in cells that had morphology of intermediate neural progenitors and immature granule neurons showing that these cells remained preserved with aging. PSA-NCAM: Polysialylated neural cell adhesion molecule.

endorphin is a posttranslational cleavage product of proopiomelanocortin [173], Paul and colleagues examined the function of proopiomelanocortin-expressing neurons in the neural stem cell micro-environment. Using state-of-the-art techniques including sophisticated *in vivo* inducible genetic approaches, such as lineage-tracing Cre/loxP mediated technologies in combination with viral vectors, the authors increased acutely proopiomelanocortin-expressing neurons activity using activating DREADDs (Designer Receptors Exclusively Activated by Designer Drugs) or selectively eliminated these neurons. These experiments revealed that proopiomelanocortin-expressing neurons affect neural stem cell proliferation in the anterior ventral subventricular zone [171,172]. Importantly, Paul and colleagues analyzed and manipulated hypothalamic neural activity in mice fed *ad libitum* or fasted, indicating that hunger and satiety states specifically regulate neural stem cells proliferation in the anterior ventral subventricular zone through proopiomelanocortin-expressing axons. This new study reveals a new constituent of the neural stem cell niche and shows that distal brain activity may regulate subventricular zone neural stem cell behavior.

## 7. Clinical relevance

Rodent models aim to recreate features of human physiology as closely as possible. Nevertheless, whether the same phenomena that is observed in mouse models occur in humans needs to be questioned. Also, whether there is conservation in the cellular and molecular mechanisms of regulation of neural stem cells in humans and mice remains poorly explored.

Nearly all our understanding of the neural stem cell behavior derives from studies in rodent models, and our knowledge about neural stem cells in the human brain is still very limited. Comparative analyses of adult neurogenesis have uncovered a big variety in this phenomenon among different species [174]. Neuroanatomical experiments and

modern techniques, such as radiocarbon dating, have proved that neural stem cells are present in the adult human brain [175]. The subventricular zone neurogenic niche differs between mice and humans, based on the cell types that form this area [176]. Newly formed neural progenitors in this zone also differ in their fate, becoming medium spiny neurons in the striatum [177], instead of forming olfactory interneurons as in mice [178]. Recent also have also shown that neural stem cells are present in the hypothalamic neurogenic region in the human adult brain [179,180].

Although some studies have suggested that hippocampal neural stem cell niche in humans resembles the one in rodents, and the fate of neural stem cells are dentate granule neurons in both species [181,182], a recent intriguing study in *Nature* has challenged this concept [183]. Sorrells and colleagues found that the number of hippocampal neural stem cells and young neurons decreases in the first one year of age, and only few isolated young neurons are present in the hippocampus in the first decade of life [183]. Strikingly, the authors did not detect any neural stem cells or young neurons in the adult human dentate gyrus [183]. Additionally, Sorrells and colleagues assessed autopsies hippocampi from monkeys. Although the authors detected neurogenesis in the early postnatal life, it diminished with aging [183] (Fig. 5). In contrast, another study published in *Cell Stem Cell* at the same time suggested just the opposite [184]. Boldrini and colleagues analyzed the hippocampi of healthy humans of different ages. The authors revealed that neurogenesis in healthy older individuals without any neurological dysfunction was preserved with aging [184] (Fig. 6). The discrepancies between these two studies may be due to several technical issues, such as limitations of specific neural stem cell markers and quantitative aspects in humans, as it was elegantly discussed in a recent mini-review [185]. Very little is known about the niches of the neural stem cells in the distinct adult human brain neurogenic areas.



## 8. Conclusions and perspectives

The works discussed in this review illustrate the complexity of the microenvironments where neural stem cells are located in the brain. Multiple cell populations contribute to the complex maintenance and regulation of neural stem cells. The availability of refined genetic technologies has demonstrated that changes to the niche content may have profound effects on neural stem cell behavior. Recombination-based technologies provide powerful ways to interrogate the cellular and molecular components of the neural stem cell niches. Future clarification of the interactions between neural stem cells and their microenvironments in pathological conditions may lead to improved methods to exploit the clinical potential of neural stem cells. In the future, targeting the niche itself could become an attractive potential alternative for the treatment of neurological illnesses. The balance of extrinsic effects from the neurogenic niche can also differ under distinct physiological circumstances. Whether newborn, adult, and aged neural stem cells have variable physiological demands remains poorly understood. The examination of how the neural stem cells niches age will reveal essential information for the treatment of age-related neurological disorders. The biggest challenge for the future still will be to translate animal research into humans. Enhancing the availability of human brain tissue samples will be fundamental to reach this aim. There is also a need for novel strategies to study the generation of new neurons in vivo in the adult human brain. A more detailed analysis of single cell phenotypes in the neurogenic niches, by for instance single-neural stem cell RNA sequencing as well as by single-cell RNA sequencing of each of the neural stem cell niche components in the adult brain, will provide beneficial knowledge. Immense progress has been made in our understanding of the importance and the complexity of the niche to the function of neural stem cells and to the physiology of the organism as a whole. Nevertheless, the best is yet to come.

## Disclosures

The authors indicate no potential conflicts of interest.

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## References

- [1] J.W. Truman, Developmental neuroethology of insect metamorphosis, *J. Neurobiol.* 23 (10) (1992) 1404–1422.
- [2] A. Alvarez-Buylla, J.R. Kirn, F. Nottebohm, Birth of projection neurons in adult avian brain may be related to perceptual or motor learning, *Science* 249 (4975) (1990) 1444–1446.
- [3] A. Kriegstein, A. Alvarez-Buylla, The glial nature of embryonic and adult neural stem cells, *Annu. Rev. Neurosci.* 32 (2009) 149–184.
- [4] B.A. Reynolds, S. Weiss, Generation of neurons and astrocytes from isolated cells of the adult mammalian central nervous system, *Science* 255 (5052) (1992) 1707–1710.
- [5] L. Li, H. Clevers, Coexistence of quiescent and active adult stem cells in mammals, *Science* 327 (5965) (2010) 542–545.
- [6] P.P. Naik, A. Birbrair, S.K. Bhutia, Mitophagy-driven metabolic switch reprograms stem cell fate, *Cell. Mol. Life Sci.*: CMLS (2018).
- [7] A. Arvidsson, T. Collin, D. Kirik, Z. Kokaia, O. Lindvall, Neuronal replacement from endogenous precursors in the adult brain after stroke, *Nat. Med.* 8 (9) (2002) 963–970.
- [8] R. Lin, L. Iacovitti, Classic and novel stem cell niches in brain homeostasis and repair, *Brain Res.* 1628 (Pt B) (2015) 327–342.
- [9] M.V. Kokoeva, H. Yin, J.S. Flier, Evidence for constitutive neural cell proliferation in the adult murine hypothalamus, *J. Comp. Neurol.* 505 (2) (2007) 209–220.
- [10] A.A. Pierce, A.W. Xu, De novo neurogenesis in adult hypothalamus as a compensatory mechanism to regulate energy balance, *J. Neurosci.* 30 (2) (2010) 723–730.
- [11] D.E. McNay, N. Briancon, M.V. Kokoeva, E. Maratos-Flier, J.S. Flier, Remodeling of the arcuate nucleus energy-balance circuit is inhibited in obese mice, *J. Clin. Invest.* 122 (1) (2012) 142–152.
- [12] F. Calzolari, J. Michel, E.V. Baumgart, F. Theis, M. Gotz, J. Ninkovic, Fast clonal expansion and limited neural stem cell self-renewal in the adult subependymal zone, *Nat. Neurosci.* 18 (4) (2015) 490–492.
- [13] A.M. Bond, G.L. Ming, H. Song, Adult mammalian neural stem cells and neurogenesis: five decades later, *Cell Stem Cell* 17 (4) (2015) 385–395.
- [14] R.A. Ihrie, A. Alvarez-Buylla, Lake-front property: a unique germinal niche by the lateral ventricles of the adult brain, *Neuron* 70 (4) (2011) 674–686.
- [15] B. Seri, J.M. Garcia-Verdugo, B.S. McEwen, A. Alvarez-Buylla, Astrocytes give rise to new neurons in the adult mammalian hippocampus, *J. Neurosci.* 21 (18) (2001) 7153–7160.
- [16] F.H. Gage, S. Temple, Neural stem cells: generating and regenerating the brain, *Neuron* 80 (3) (2013) 588–601.
- [17] G. Kempermann, H.G. Kuhn, F.H. Gage, More hippocampal neurons in adult mice living in an enriched environment, *Nature* 386 (6624) (1997) 493–495.
- [18] H. van Praag, T. Shubert, C. Zhao, F.H. Gage, Exercise enhances learning and hippocampal neurogenesis in aged mice, *J. Neurosci.* 25 (38) (2005) 8680–8685.
- [19] E. Drapeau, W. Mayo, C. Aurousseau, M. Le Moal, P.V. Piazza, D.N. Abrous, Spatial memory performances of aged rats in the water maze predict levels of hippocampal neurogenesis, *Proc. Natl. Acad. Sci. U. S. A.* 100 (24) (2003) 14385–14390.
- [20] A. Alvarez-Buylla, D.G. Herrera, H. Wichterle, The subventricular zone: source of neuronal precursors for brain repair, *Prog. Brain Res.* 127 (2000) 1–11.
- [21] V. Breton-Provencher, M. Lemasson, M.R. Peralta 3rd, A. Saghatelian, Interneurons during adulthood are required for the normal functioning of the olfactory bulb network and for the execution of selected olfactory behaviors, *J. Neurosci.* 29 (48) (2009) 15245–15257.
- [22] G. Gheusi, H. Cremer, H. McLean, G. Chazal, J.D. Vincent, P.M. Lledo, Importance of newly generated neurons in the adult olfactory bulb for odor discrimination, *Proc. Natl. Acad. Sci. U. S. A.* 97 (4) (2000) 1823–1828.
- [23] M.A. Bonaguidi, M.A. Wheeler, J.S. Shapiro, R.P. Stadel, G.J. Sun, G.L. Ming, H. Song, In vivo clonal analysis reveals self-renewing and multipotent adult neural stem cell characteristics, *Cell* 145 (7) (2011) 1142–1155.
- [24] Z. Mirzadeh, F.T. Merkle, M. Soriano-Navarro, J.M. Garcia-Verdugo, A. Alvarez-Buylla, Neural stem cells confer unique pinwheel architecture to the ventricular surface in neurogenic regions of the adult brain, *Cell Stem Cell* 3 (3) (2008) 265–278.
- [25] D. Bottai, G. Scesa, D. Cigognini, R. Adami, E. Nicora, S. Abrignani, A.M. Di Giulio, A. Gorio, Third trimester NG2-positive amniotic fluid cells are effective in improving repair in spinal cord injury, *Exp. Neurol.* 254 (2014) 121–133.
- [26] J.P. Andreotti, A.E. Paiva, P. Prazeres, D.A.P. Guerra, W.N. Silva, R.S. Vaz, A. Mintz, A. Birbrair, The role of natural killer cells in the uterine microenvironment during pregnancy, *Cell. Mol. Immunol.* (2018).
- [27] P.O. Azevedo, A.E. Paiva, G.S.P. Santos, L. Lousado, J.P. Andreotti, I.F.G. Sena, A. Mintz, A. Birbrair, Cross-talk between lung cancer and bones results in neurotrophils that promote tumor progression, *Cancer Metastasis Rev.* (2018).
- [28] D.A.P. Guerra, A.E. Paiva, I.F.G. Sena, P.O. Azevedo, M.L. Batista Jr., A. Mintz, A. Birbrair, Adipocytes role in the bone marrow niche, *Cytomet. Part A: J. Int. Soc. Anal. Cytol.* 93 (2) (2018) 167–171.
- [29] A.E. Paiva, L. Lousado, V.M. Almeida, J.P. Andreotti, G.S.P. Santos, P.O. Azevedo, I.F.G. Sena, P. Prazeres, I.T. Borges, V. Azevedo, A. Mintz, A. Birbrair, Endothelial cells as precursors for osteoblasts in the metastatic prostate cancer bone, *Neoplasia* 19 (11) (2017) 928–931.
- [30] L. Lousado, P. Prazeres, J.P. Andreotti, A.E. Paiva, P.O. Azevedo, G.S.P. Santos, R. Filev, A. Mintz, A. Birbrair, Schwann cell precursors as a source for adrenal gland chromaffin cells, *Cell Death Dis.* 8 (10) (2017) e3072.
- [31] A.C.Z. Rodrigues, M.L. Messi, Z.M. Wang, M.C. Abba, A. Pereyra, A. Birbrair, T. Zhang, M. O'Meara, P. Kwan, E.I.S. Lopez, M.S. Willis, A. Mintz, D.C. Files, C. Furdul, R.W. Oppenheim, O. Delbono, The sympathetic nervous system regulates skeletal muscle motor innervation and acetylcholine receptor stability, *Acta Physiol.* (2018) e13195.
- [32] A. Birbrair, Stem cell microenvironments and beyond, *Adv. Exp. Med. Biol.* 1041 (2017) 1–3.
- [33] G.L. Ming, H. Song, Adult neurogenesis in the mammalian brain: significant answers and significant questions, *Neuron* 70 (4) (2011) 687–702.
- [34] I. Decimo, F. Bifari, M. Krampera, G. Fumagalli, Neural stem cell niches in health and diseases, *Curr. Pharm. Des.* 18 (13) (2012) 1755–1783.
- [35] C. Goritz, J. Frisen, Neural stem cells and neurogenesis in the adult, *Cell Stem Cell* 10 (6) (2012) 657–659.
- [36] F.T. Merkle, L.C. Fuentealba, T.A. Sanders, L. Magno, N. Kassaris, A. Alvarez-Buylla, Adult neural stem cells in distinct microdomains generate previously unknown interneuron types, *Nat. Neurosci.* 17 (2) (2014) 207–214.
- [37] R. Fiorelli, K. Azim, B. Fischer, O. Raineteau, Adding a spatial dimension to postnatal ventricular-subventricular zone neurogenesis, *Development* 142 (12) (2015) 2109–2120.
- [38] L.C. Fuentealba, S.B. Rompani, J.I. Parraguez, K. Obner, R. Romero, C.L. Cepko,

- A. Alvarez-Buylla, Embryonic origin of postnatal neural stem cells, *Cell* 161 (7) (2015) 1644–1655.
- [39] F.T. Merkle, Z. Mirzadeh, A. Alvarez-Buylla, Mosaic organization of neural stem cells in the adult brain, *Science* 317 (5836) (2007) 381–384.
- [40] A. Isomura, R. Kageyama, Ultradian oscillations and pulses: coordinating cellular responses and cell fate decisions, *Development* 141 (19) (2014) 3627–3636.
- [41] M. Kohwi, C.Q. Doe, Temporal fate specification and neural progenitor competence during development, *Nature reviews, Neuroscience* 14 (12) (2013) 823–838.
- [42] A.D. Ramos, R.E. Andersen, S.J. Liu, T.J. Nowakowski, S.J. Hong, C. Gertz, R.D. Salinas, H. Zarabi, A.R. Kriegstein, D.A. Lim, The long noncoding RNA Pnky regulates neuronal differentiation of embryonic and postnatal neural stem cells, *Cell Stem Cell* 16 (4) (2015) 439–447.
- [43] A. Capela, S. Temple, LeX/ssea-1 is expressed by adult mouse CNS stem cells, identifying them as nonependymal, *Neuron* 35 (5) (2002) 865–875.
- [44] S. Corti, M. Nizzardo, M. Nardini, C. Donadoni, F. Locatelli, D. Papadimitriou, S. Salani, R. Del Bo, S. Ghezzi, S. Strazzer, N. Bresolin, G.P. Comi, Isolation and characterization of murine neural stem/progenitor cells based on Prominin-1 expression, *Exp. Neurol.* 205 (2) (2007) 547–562.
- [45] P. Barraud, L. Thompson, D. Kirik, A. Bjorklund, M. Parmar, Isolation and characterization of neural precursor cells from the Sox1-GFP reporter mouse, *Eur. J. Neurosci.* 22 (7) (2005) 1555–1569.
- [46] A. Kawaguchi, T. Miyata, K. Sawamoto, N. Takashita, A. Murayama, W. Akamatsu, M. Ogawa, M. Okabe, Y. Tano, S.A. Goldman, H. Okano, Nestin-EGFP transgenic mice: visualization of the self-renewal and multipotency of CNS stem cells, *Mol. Cell. Neurosci.* 17 (2) (2001) 259–273.
- [47] A. Birbrair, A. Sattiraju, D. Zhu, G. Zulato, I. Batista, V.T. Nguyen, M.L. Messi, K.K. Solingapuram Sai, F.C. Marini, O. Delbono, A. Mintz, Novel peripherally derived neural-like stem cells as therapeutic carriers for treating glioblastomas, *Stem Cells Transl. Med.* 6 (2) (2017) 471–481.
- [48] A. Birbrair, Z.M. Wang, M.L. Messi, G.N. Enikolopov, O. Delbono, Nestin-GFP transgene reveals neural precursor cells in adult skeletal muscle, *PLoS One* 6 (2) (2011) e16816.
- [49] F. Doetsch, L. Petreanu, I. Caille, J.M. Garcia-Verdugo, A. Alvarez-Buylla, EGF converts transit-amplifying neurogenic precursors in the adult brain into multipotent stem cells, *Neuron* 36 (6) (2002) 1021–1034.
- [50] Y. Luo, V. Coskun, A. Liang, J. Yu, L. Cheng, W. Ge, Z. Shi, K. Zhang, C. Li, Y. Cui, H. Lin, D. Luo, J. Wang, C. Lin, Z. Dai, H. Zhu, J. Zhang, J. Liu, H. Liu, J. deVellis, S. Horvath, Y.E. Sun, S. Li, Single-cell transcriptome analyses reveal signals to activate dormant neural stem cells, *Cell* 161 (5) (2015) 1175–1186.
- [51] E. Pastrana, L.C. Cheng, F. Doetsch, Simultaneous prospective purification of adult subventricular zone neural stem cells and their progeny, *Proc. Natl. Acad. Sci. U. S. A.* 106 (15) (2009) 6387–6392.
- [52] P. Codega, V. Silva-Vargas, A. Paul, A.R. Maldonado-Soto, A.M. Deleo, E. Pastrana, F. Doetsch, Prospective identification and purification of quiescent adult neural stem cells from their in vivo niche, *Neuron* 82 (3) (2014) 545–559.
- [53] E. Llorens-Bobadilla, S. Zhao, A. Baser, G. Saiz-Castro, K. Zwadlo, A. Martin-Villalba, Single-cell transcriptomics reveals a population of dormant neural stem cells that become activated upon brain injury, *Cell Stem Cell* 17 (3) (2015) 329–340.
- [54] K. Azim, S. Zweifel, F. Klaus, K. Yoshikawa, I. Amrein, O. Raineteau, Early decline in progenitor diversity in the marmoset lateral ventricle, *Cereb. Cortex* 23 (4) (2013) 922–931.
- [55] N.A. DeCarolis, M. Mechanic, D. Petrik, A. Carlton, J.L. Ables, S. Malhotra, R. Bachoo, M. Gotz, D.C. Lagace, A.J. Eisch, In vivo contribution of nestin- and GLAST-lineage cells to adult hippocampal neurogenesis, *Hippocampus* 23 (8) (2013) 708–719.
- [56] E. Gebara, M.A. Bonaguidi, R. Beckervordersandforth, S. Sultan, F. Udry, P.J. Gijs, D.C. Lie, G.L. Ming, H. Song, N. Toni, Heterogeneity of radial glia-like cells in the adult Hippocampus, *Stem Cells* 34 (4) (2016) 997–1010.
- [57] D.G. Amaral, M.P. Witter, The three-dimensional organization of the hippocampal formation: a review of anatomical data, *Neuroscience* 31 (3) (1989) 571–591.
- [58] M.V. Wu, A. Sahay, R.S. Duman, R. Hen, Functional differentiation of adult-born neurons along the septotemporal axis of the dentate gyrus, *Cold Spring Harb. Perspect. Biol.* 7 (8) (2015) a018978.
- [59] J. Sun, M.A. Bonaguidi, H. Jun, J.U. Guo, G.J. Sun, B. Will, Z. Yang, M.H. Jang, H. Song, G.L. Ming, K.M. Christian, A septo-temporal molecular gradient of sfrp3 in the dentate gyrus differentially regulates quiescent adult hippocampal neural stem cell activation, *Mol. Brain* 8 (2015) 52.
- [60] M.H. Jang, M.A. Bonaguidi, Y. Kitabatake, J. Sun, J. Song, E. Kang, H. Jun, C. Zhong, Y. Su, J.U. Guo, M.X. Wang, K.A. Sailor, J.Y. Kim, Y. Gao, K.M. Christian, G.L. Ming, H. Song, Secreted frizzled-related protein 3 regulates activity-dependent adult hippocampal neurogenesis, *Cell Stem Cell* 12 (2) (2013) 215–223.
- [61] V.C. Piatti, M.G. Davies-Sala, M.S. Esposito, L.A. Mongiat, M.F. Trincherio, A.F. Schinder, The timing for neuronal maturation in the adult hippocampus is modulated by local network activity, *J. Neurosci.* 31 (21) (2011) 7715–7728.
- [62] S. Jinno, Topographic differences in adult neurogenesis in the mouse hippocampus: a stereology-based study using endogenous markers, *Hippocampus* 21 (5) (2011) 467–480.
- [63] J. Ninkovic, M. Gotz, How to make neurons—thoughts on the molecular logic of neurogenesis in the central nervous system, *Cell Tissue Res.* 359 (1) (2015) 5–16.
- [64] M.K. Lehtinen, C.S. Bjornsson, S.M. Dymecki, R.J. Gilbertson, D.M. Holtzman, E.S. Monuki, The choroid plexus and cerebrospinal fluid: emerging roles in development, disease, and therapy, *J. Neurosci.* 33 (45) (2013) 17553–9.
- [65] Z.B. Redzic, J.E. Preston, J.A. Duncan, A. Chodobski, J. Szymdynger-Chodobska, The choroid plexus-cerebrospinal fluid system: from development to aging, *Curr. Top. Dev. Biol.* 71 (2005) 1–52.
- [66] C.E. Johanson, J.A. Duncan 3rd, P.M. Klinge, T. Brinker, E.G. Stopa, G.D. Silverberg, Multiplicity of cerebrospinal fluid functions: new challenges in health and disease, *Cerebrospinal Fluid Res.* 5 (2008) 10.
- [67] M.K. Lehtinen, M.W. Zappaterra, X. Chen, Y.J. Yang, A.D. Hill, M. Lun, T. Maynard, D. Gonzalez, S. Kim, P. Ye, A.J. D’Ercole, E.T. Wong, A.S. LaMantia, C.A. Walsh, The cerebrospinal fluid provides a proliferative niche for neural progenitor cells, *Neuron* 69 (5) (2011) 893–905.
- [68] V. Silva-Vargas, A.R. Maldonado-Soto, D. Mizrak, P. Codega, F. Doetsch, Age-dependent niche signals from the choroid plexus regulate adult neural stem cells, *Cell Stem Cell* 19 (5) (2016) 643–652.
- [69] K. Sawamoto, H. Wichterle, O. Gonzalez-Perez, J.A. Cholfin, M. Yamada, N. Spassky, N.S. Murcia, J.M. Garcia-Verdugo, O. Marin, J.L. Rubenstein, M. Tessier-Lavigne, H. Okano, A. Alvarez-Buylla, New neurons follow the flow of cerebrospinal fluid in the adult brain, *Science* 311 (5761) (2006) 629–632.
- [70] P.A. Johansson, M. Irmeler, D. Acampora, J. Beckers, A. Simeone, M. Gotz, The transcription factor Otx2 regulates choroid plexus development and function, *Development* 140 (5) (2013) 1055–1066.
- [71] P.A. Johansson, S. Cappello, M. Gotz, Stem cells niches during development—lessons from the cerebral cortex, *Curr. Opin. Neurobiol.* 20 (4) (2010) 400–407.
- [72] A.M. Falcao, F. Marques, A. Novais, N. Sousa, J.A. Palha, J.C. Sousa, The path from the choroid plexus to the subventricular zone: go with the flow!, *Front. Cell. Neurosci.* 6 (2012) 34.
- [73] H.B. Stolp, Neuroepoietic cytokines in normal brain development and neurodevelopmental disorders, *Mol. Cell. Neurosci.* 53 (2013) 63–68.
- [74] K.M. Dziegielewska, G.W. Knott, N.R. Saunders, The nature and composition of the internal environment of the developing brain, *Cell. Mol. Neurobiol.* 20 (1) (2000) 41–56.
- [75] A.C. Delgado, S.R. Ferron, D. Vicente, E. Porlan, A. Perez-Villalba, C.M. Trujillo, P. D’Ocon, I. Farinas, Endothelial NT-3 delivered by vasculature and CSF promotes quiescence of subependymal neural stem cells through nitric oxide induction, *Neuron* 83 (3) (2014) 572–585.
- [76] M.W. Zappaterra, M.K. Lehtinen, The cerebrospinal fluid: regulator of neurogenesis, behavior, and beyond, *Cell. Mol. Life Sci.* CMLS 69 (17) (2012) 2863–2878.
- [77] D. Petrik, M.H. Myoga, S. Grade, N.J. Gerkau, M. Pusch, C.R. Rose, B. Grothe, M. Gotz, Epithelial sodium channel regulates adult neural stem cell proliferation in a flow-dependent manner, *Cell Stem Cell* 22 (6) (2018) 865–878 e8.
- [78] A. Birbrair, I.D.T. Borges, I.F. Gilson Sena, G.G. Almeida, L. da Silva Meirelles, R. Goncalves, A. Mintz, O. Delbono, How plastic are pericytes? *Stem Cells Dev.* 26 (14) (2017) 1013–1019.
- [79] D.O. Dias, H. Kim, D. Holl, B. Werne Solnestam, J. Lundeberg, M. Carlen, C. Goritz, J. Frisen, Reducing pericyte-derived scarring promotes recovery after spinal cord injury, *Cell* 173 (1) (2018) 153–165 e22.
- [80] Y. Zhou, A.M. Bond, J.E. Shade, Y. Zhu, C.O. Davis, X. Wang, Y. Su, K.J. Yoon, A.T. Phan, W.J. Chen, J.H. Oh, N. Marsh-Armstrong, K. Atabai, G.L. Ming, H. Song, Autocrine Mfge8 signaling prevents developmental exhaustion of the adult neural stem cell pool, *Cell Stem Cell* 23 (3) (2018) 444–452 e4.
- [81] J. Shin, D.A. Berg, Y. Zhu, J.Y. Shin, J. Song, M.A. Bonaguidi, G. Enikolopov, D.W. Nauen, K.M. Christian, G.L. Ming, H. Song, Single-cell RNA-Seq with water-fall reveals molecular cascades underlying adult neurogenesis, *Cell Stem Cell* 17 (3) (2015) 360–372.
- [82] R.K. Schneider, A. Mullally, A. Dugourd, F. Peisker, R. Hoogenboezem, P.M.H. Van Strien, E.M. Bindels, D. Heckl, G. Busche, D. Fleck, G. Muller-Newen, J. Wongboonsin, M. Ventura Ferreira, V.G. Puelles, J. Saez-Rodriguez, B.L. Ebert, B.D. Humphreys, R. Kramann, Gli1+ mesenchymal stromal cells are a key driver of bone marrow fibrosis and an important cellular therapeutic target, *Cell Stem Cell* 20 (6) (2017) 785–800 e8.
- [83] I.F.G. Sena, P. Prazeres, G.S.P. Santos, I.T. Borges, P.O. Azevedo, J.P. Andreotti, V.M. Almeida, A.E. Paiva, D.A.P. Guerra, L. Lousado, L. Souto, A. Mintz, A. Birbrair, Identity of Gli1+ cells in the bone marrow, *Exp. Hematol.* 54 (2017) 12–16.
- [84] I.F.G. Sena, I.T. Borges, L. Lousado, P.O. Azevedo, J.P. Andreotti, V.M. Almeida, A.E. Paiva, G.S.P. Santos, D.A.P. Guerra, P. Prazeres, L. Souto, A. Mintz, A. Birbrair, LepR+ cells dispute hegemony with Gli1+ cells in bone marrow fibrosis, *Cell Cycle* (2017) 1–5.
- [85] S. Yoshida, M. Sueno, Y. Nabeshima, A vasculature-associated niche for undifferentiated spermatogonia in the mouse testis, *Science* 317 (5845) (2007) 1722–1726.
- [86] W. Tang, D. Zeve, J.M. Suh, D. Bosnakovski, M. Kyba, R.E. Hammer, M.D. Tallquist, J.M. Graff, White fat progenitor cells reside in the adipose vasculature, *Science* 322 (5901) (2008) 583–586.
- [87] N. Asada, Y. Kunisaki, H. Pierce, Z. Wang, N.F. Fernandez, A. Birbrair, A. Ma’ayan, P.S. Frenette, Differential cytokine contributions of perivascular haematopoietic stem cell niches, *Nat. Cell Biol.* 19 (3) (2017) 214–223.
- [88] J.A. Khan, A. Mendelson, Y. Kunisaki, A. Birbrair, Y. Kou, A. Arnal-Estape, S. Pinho, P. Ciero, F. Nakahara, A. Ma’ayan, A. Bergman, M. Merad, P.S. Frenette, Fetal liver hematopoietic stem cell niches associate with portal vessels, *Science* 351 (6269) (2016) 176–180.
- [89] T.D. Palmer, A.R. Willhoite, F.H. Gage, Vascular niche for adult hippocampal neurogenesis, *J. Comp. Neurol.* 425 (4) (2000) 479–494.
- [90] Q. Shen, Y. Wang, E. Kokovay, G. Lin, S.M. Chuang, S.K. Goderie, B. Roysam, S. Temple, Adult SVZ stem cells lie in a vascular niche: a quantitative analysis of niche cell-cell interactions, *Cell Stem Cell* 3 (3) (2008) 289–300.
- [91] M. Tavazoie, L. Van der Veken, V. Silva-Vargas, M. Louissaint, L. Colonna, B. Zaidi, J.M. Garcia-Verdugo, F. Doetsch, A specialized vascular niche for adult neural stem cells, *Cell Stem Cell* 3 (3) (2008) 279–288.

- [92] J.C. Culver, T.J. Vadakkan, M.E. Dickinson, A specialized microvascular domain in the mouse neural stem cell niche, *PLoS One* 8 (1) (2013) e53546.
- [93] I. Kazanis, J.D. Lathia, T.J. Vadakkan, E. Raborn, R. Wan, M.R. Mughal, D.M. Eckley, T. Sasaki, B. Patton, M.P. Mattson, K.K. Hirschi, M.E. Dickinson, C. French-Constant, Quiescence and activation of stem and precursor cell populations in the subependymal zone of the mammalian brain are associated with distinct cellular and extracellular matrix signals, *J. Neurosci.* 30 (29) (2010) 9771–9781.
- [94] B. Lacar, S.Z. Young, J.C. Platel, A. Bordey, Gap junction-mediated calcium waves define communication networks among murine postnatal neural progenitor cells, *Eur. J. Neurosci.* 34 (12) (2011) 1895–1905.
- [95] D. Petrik, S. Yun, S.E. Latchney, S. Kamrudin, J.A. LeBlanc, J.A. Bibb, A.J. Eisch, Early postnatal *in vivo* gliogenesis from nestin-lineage progenitors requires *cdk5*, *PLoS One* 8 (8) (2013) e72819.
- [96] J.L. Mignone, V. Kukekov, A.S. Chiang, D. Steindler, G. Enikolopov, Neural stem and progenitor cells in nestin-GFP transgenic mice, *J. Comp. Neurol.* 469 (3) (2004) 311–324.
- [97] E. Kokovay, S. Goderie, Y. Wang, S. Lotz, G. Lin, Y. Sun, B. Roysam, Q. Shen, S. Temple, Adult SVZ lineage cells home to and leave the vascular niche via differential responses to SDF1/CXCR4 signaling, *Cell Stem Cell* 7 (2) (2010) 163–173.
- [98] L. Cao, X. Jiao, D.S. Zuzga, Y. Liu, D.M. Fong, D. Young, M.J. During, VEGF links hippocampal activity with neurogenesis, learning and memory, *Nat. Genet.* 36 (8) (2004) 827–835.
- [99] K. Jin, Y. Zhu, Y. Sun, X.O. Mao, L. Xie, D.A. Greenberg, Vascular endothelial growth factor (VEGF) stimulates neurogenesis *in vitro* and *in vivo*, *Proc. Natl. Acad. Sci. U. S. A.* 99 (18) (2002) 11946–11950.
- [100] T. Licht, I. Goshen, A. Avital, T. Kreisel, S. Zuber, R. Eavri, M. Segal, R. Yirmiya, E. Keshet, Reversible modulations of neuronal plasticity by VEGF, *Proc. Natl. Acad. Sci. U. S. A.* 108 (12) (2011) 5081–5086.
- [101] H. Udo, Y. Yoshida, T. Kino, K. Ohnuki, W. Mizunoya, T. Mukuda, H. Sugiyama, Enhanced adult neurogenesis and angiogenesis and altered affective behaviors in mice overexpressing vascular endothelial growth factor 120, *J. Neurosci.* 28 (53) (2008) 14522–14536.
- [102] A.C. Pereira, D.E. Huddleston, A.M. Brickman, A.A. Sosunov, R. Hen, G.M. McKhann, R. Sloan, F.H. Gage, T.R. Brown, S.A. Small, An *in vivo* correlate of exercise-induced neurogenesis in the adult dentate gyrus, *Proc. Natl. Acad. Sci. U. S. A.* 104 (13) (2007) 5638–5643.
- [103] K. Van der Borght, D.E. Kobor-Nyakas, K. Klauke, B.J. Eggen, C. Nyakas, E.A. Van der Zee, P. Meerlo, Physical exercise leads to rapid adaptations in hippocampal vasculature: temporal dynamics and relationship to cell proliferation and neurogenesis, *Hippocampus* 19 (10) (2009) 928–936.
- [104] C. Ottone, B. Krusche, A. Whitby, M. Clements, G. Quadrato, M.E. Pitulescu, R.H. Adams, S. Parrinello, Direct cell-cell contact with the vascular niche maintains quiescent neural stem cells, *Nat. Cell Biol.* 16 (11) (2014) 1045–1056.
- [105] Y. Sato, Y. Uchida, J. Hu, T.L. Young-Pearse, T. Niikura, Y.S. Mukoyama, Soluble APP functions as a vascular niche signal that controls adult neural stem cell number, *Development* (2017).
- [106] P.O. Azevedo, L. Lousado, A.E. Paiva, J.P. Andreotti, G.S.P. Santos, I.F.G. Sena, P. Prazeres, R. Filev, A. Mintz, A. Birbrair, Endothelial cells maintain neural stem cells quiescent in their niche, *Neuroscience* 363 (2017) 62–65.
- [107] F. Arai, A. Hirao, M. Ohmura, H. Sato, S. Matsuoka, K. Takubo, K. Ito, G.Y. Koh, T. Suda, Tie2/angiopoietin-1 signaling regulates hematopoietic stem cell quiescence in the bone marrow niche, *Cell* 118 (2) (2004) 149–161.
- [108] N. Takakura, X.L. Huang, T. Naruse, I. Hamaguchi, D.J. Dumont, G.D. Yancopoulos, T. Suda, Critical role of the Tie2 endothelial cell receptor in the development of definitive hematopoiesis, *Immunity* 9 (5) (1998) 677–686.
- [109] D.Y. Park, J. Lee, J. Kim, K. Kim, S. Hong, S. Han, Y. Kubota, H.G. Augustin, L. Ding, J.W. Kim, H. Kim, Y. He, R.H. Adams, G.Y. Koh, Plastic roles of pericytes in the blood-retinal barrier, *Nat. Commun.* 8 (2017) 15296.
- [110] A. Mishra, J.P. Reynolds, Y. Chen, A.V. Gourine, D.A. Rusakov, D. Attwell, Astrocytes mediate neurovascular signaling to capillary pericytes but not to arterioles, *Nat. Neurosci.* 19 (12) (2016) 1619–1627.
- [111] K. Kisler, A.R. Nelson, A. Montagne, B.V. Zlokovic, Cerebral blood flow regulation and neurovascular dysfunction in Alzheimer disease, *Nat. Rev. Neurosci.* 18 (7) (2017) 419–434.
- [112] I. Bechmann, J. Priller, A. Kovac, M. Bontert, T. Wehner, F.F. Klett, J. Bohsung, M. Stuschke, U. Dirnagl, R. Nitsch, Immune surveillance of mouse brain perivascular spaces by blood-borne macrophages, *Eur. J. Neurosci.* 14 (10) (2001) 1651–1658.
- [113] W.N. Silva, P. Prazeres, A.E. Paiva, L. Lousado, A.O.M. Turqueti, R.S.N. Barreto, E.C. de Alvarenga, M.A. Miglino, R. Goncalves, A. Mintz, A. Birbrair, Macrophage-derived GPNMB accelerates skin healing, *Exp. Dermatol.* (2018).
- [114] P. Prazeres, V.M. Almeida, L. Lousado, J.P. Andreotti, A.E. Paiva, G.S.P. Santos, P.O. Azevedo, L. Souto, G.G. Almeida, R. Filev, A. Mintz, R. Goncalves, A. Birbrair, Macrophages generate pericytes in the developing brain, *Cell. Mol. Neurobiol.* 38 (4) (2018) 777–782.
- [115] M. Crisan, M. Corselli, W.C. Chen, B. Peault, Perivascular cells for regenerative medicine, *J. Cell. Mol. Med.* (2012).
- [116] C. Soderblom, X. Luo, E. Blumenthal, E. Bray, K. Lyapichev, J. Ramos, V. Krishnan, C. Lai-Hsu, K.K. Park, P. Tsoulfas, J.K. Lee, Perivascular fibroblasts form the fibrotic scar after contusive spinal cord injury, *J. Neurosci.* 33 (34) (2013) 13882–13887.
- [117] G.J. Guillemain, B.J. Brew, Microglia, macrophages, perivascular macrophages, and pericytes: a review of function and identification, *J. Leukoc. Biol.* 75 (3) (2004) 388–397.
- [118] M. Wanjare, S. Kusuma, S. Gerecht, Perivascular cells in blood vessel regeneration, *Biotechnol. J.* 8 (4) (2013) 434–447.
- [119] A. Birbrair, T. Zhang, Z.M. Wang, M.L. Messi, J.D. Olson, A. Mintz, O. Delbono, Type-2 pericytes participate in normal and tumoral angiogenesis, *Am. J. Physiol. Cell Physiol.* 307 (1) (2014) C25–38.
- [120] G.S.P. Santos, L.A.V. Magno, M.A. Romano-Silva, A. Mintz, A. Birbrair, Pericytes plasticity in the brain, *Neurosci. Bull.* (2018).
- [121] V.M. Almeida, A.E. Paiva, I.F.G. Sena, A. Mintz, L.A.V. Magno, A. Birbrair, Pericytes make spinal cord breathless after injury, *Neuroscientist* (2017).
- [122] W.N. Silva, C. Leonel, P. Prazeres, I.F.G. Sena, D.A.P. Guerra, D. Heller, I.M.A. Diniz, V. Fortuna, A. Mintz, A. Birbrair, Role of Schwann cells in cutaneous wound healing, *Wound Repair Regen.* (2018).
- [123] P.H.D.M. Prazeres, A.O.M. Turqueti, P.O. Azevedo, R.S.N. Barreto, M.A. Miglino, A. Mintz, O. Delbono, A. Birbrair, Perivascular cell *cv* integrins as a target to treat skeletal muscle fibrosis, *Int. J. Biochem. Cell Biol.* (2018).
- [124] D.A.P. Guerra, A.E. Paiva, I.F.G. Sena, P.O. Azevedo, W.N. Silva, A. Mintz, A. Birbrair, Targeting glioblastoma-derived pericytes improves chemotherapeutic outcome, *Angiogenesis* (2018).
- [125] I.F.G. Sena, A.E. Paiva, P. Prazeres, P.O. Azevedo, L. Lousado, S.K. Bhutia, A.B. Salmina, A. Mintz, A. Birbrair, Glioblastoma-activated pericytes support tumor growth via immunosuppression, *Cancer Med.* (2018).
- [126] G.S.P. Santos, P. Prazeres, A. Mintz, A. Birbrair, Role of pericytes in the retina, *Eye* (2017).
- [127] M.A. Costa, A.E. Paiva, J.P. Andreotti, M.V. Cardoso, C.D. Cardoso, A. Mintz, A. Birbrair, Pericytes constrict blood vessels after myocardial ischemia, *J. Mol. Cell. Cardiol.* (2018).
- [128] G.C. Coatti, M. Frangini, M.C. Valadares, J.P. Gomes, N.O. Lima, N. Cavacana, A.F. Assoni, M.V. Pelatti, A. Birbrair, A.C.P. de Lima, J.M. Singer, F.M.M. Rocha, G.L. Da Silva, M.S. Mantovani, L.I. Macedo-Souza, M.F.R. Ferrari, M. Zatz, Pericytes extend survival of ALS SOD1 mice and induce the expression of anti-oxidant enzymes in the murine model and in iPSCs derived neuronal cells from an ALS patient, *Stem Cell Rev.* (2017).
- [129] P.H. Dias Moura Prazeres, I.F.G. Sena, I.D.T. Borges, P.O. de Azevedo, J.P. Andreotti, A.E. de Paiva, V.M. de Almeida, D.A. de Paula Guerra, G.S. Pinheiro Dos Santos, A. Mintz, O. Delbono, A. Birbrair, Pericytes are heterogeneous in their origin within the same tissue, *Dev. Biol.* 427 (1) (2017) 6–11.
- [130] A. Birbrair, Pericyte biology: development, Homeostasis Dis. *Adv. Exp. Med. Biol.* (2018).
- [131] A. Birbrair, P.H.D.M. Prazeres, D.C. Files, O. Delbono, Pericytes and t cells in lung injury and fibroproliferation, *Molecular and Translational Medicine book series Fibrosis in Disease*, (2019).
- [132] A. Birbrair, O. Delbono, Pericytes are essential for skeletal muscle formation, *Stem Cell Rev.* 11 (4) (2015) 547–548.
- [133] A. Birbrair, T. Zhang, Z.M. Wang, M.L. Messi, A. Mintz, O. Delbono, Pericytes at the intersection between tissue regeneration and pathology, *Clin. Sci.* 128 (2) (2015) 81–93.
- [134] A. Birbrair, T. Zhang, D.C. Files, S. Mannava, T. Smith, Z.M. Wang, M.L. Messi, A. Mintz, O. Delbono, Type-1 pericytes accumulate after tissue injury and produce collagen in an organ-dependent manner, *Stem Cell Res. Ther.* 5 (6) (2014) 122.
- [135] A. Birbrair, T. Zhang, Z.M. Wang, M.L. Messi, A. Mintz, O. Delbono, Pericytes: multitasking cells in the regeneration of injured, diseased, and aged skeletal muscle, *Front. Aging Neurosci.* 6 (2014) 245.
- [136] A. Birbrair, T. Zhang, Z.M. Wang, M.L. Messi, A. Mintz, O. Delbono, Type-1 pericytes participate in fibrous tissue deposition in aged skeletal muscle, *Am. J. Physiol. Cell Physiol.* 305 (11) (2013) C1098–113.
- [137] A. Birbrair, T. Zhang, Z.M. Wang, M.L. Messi, G.N. Enikolopov, A. Mintz, O. Delbono, Role of pericytes in skeletal muscle regeneration and fat accumulation, *Stem Cells Dev.* 22 (16) (2013) 2298–2314.
- [138] A. Birbrair, T. Zhang, Z.M. Wang, M.L. Messi, G.N. Enikolopov, A. Mintz, O. Delbono, Skeletal muscle neural progenitor cells exhibit properties of NG2-glia, *Exp. Cell Res.* 319 (1) (2013) 45–63.
- [139] A. Birbrair, T. Zhang, Z.M. Wang, M.L. Messi, G.N. Enikolopov, A. Mintz, O. Delbono, Skeletal muscle pericyte subtypes differ in their differentiation potential, *Stem Cell Res.* 10 (1) (2013) 67–84.
- [140] P.O. Azevedo, I.F.G. Sena, J.P. Andreotti, J. Carvalho-Tavares, J.C. Alves-Filho, T.M. Cunha, F.Q. Cunha, A. Mintz, A. Birbrair, Pericytes modulate myelination in the central nervous system, *J. Cell. Physiol.* (2017).
- [141] I. Borges, I. Sena, P. Azevedo, J. Andreotti, V. Almeida, A. Paiva, G. Santos, D. Guerra, P. Prazeres, L.L. Mesquita, L.S.B. Silva, C. Leonel, A. Mintz, A. Birbrair, Lung as a niche for hematopoietic progenitors, *Stem Cell Rev.* 13 (5) (2017) 567–574.
- [142] E.C. Alvarenga, W.N. Silva, R. Vasconcellos, E.J. Paredes-Gamero, A. Mintz, A. Birbrair, Promyelocytic leukemia protein in mesenchymal stem cells is essential for leukemia progression, *Ann. Hematol.* (2018).
- [143] A.E. Paiva, L. Lousado, D.A.P. Guerra, P.O. Azevedo, I.F.G. Sena, J.P. Andreotti, G.S.P. Santos, R. Goncalves, A. Mintz, A. Birbrair, Pericytes in the premetastatic niche, *Cancer Res.* (2018).
- [144] A. Birbrair, P.S. Frenette, Niche heterogeneity in the bone marrow, *Ann. N. Y. Acad. Sci.* 1370 (1) (2016) 82–96.
- [145] R. Sakuma, M. Kawahara, A. Nakano-Doi, A. Takahashi, Y. Tanaka, A. Narita, S. Kuwahara-Otani, T. Hayakawa, H. Yagi, T. Matsuyama, T. Nakagomi, Brain pericytes serve as microglia-generating multipotent vascular stem cells following ischemic stroke, *J. Neuroinflamm.* 13 (1) (2016) 57.
- [146] A. Gouveia, M. Seegobin, T.S. Kannangara, L. He, F. Wondisford, C.H. Comin, L.D.F. Costa, J.C. Beique, D.C. Lagace, B. Lacoste, J. Wang, The aPKC-CBP pathway regulates post-stroke neurovascular remodeling and functional recovery,

- Stem Cell Rep. 9 (6) (2017) 1735–1744.
- [147] T. Nakagomi, S. Kubo, A. Nakano-Doi, R. Sakuma, S. Lu, A. Narita, M. Kawahara, A. Taguchi, T. Matsuyama, Brain vascular pericytes following ischemia have multipotential stem cell activity to differentiate into neural and vascular lineage cells, *Stem Cells* 33 (6) (2015) 1962–1974.
- [148] T. Takagi, S. Yoshimura, R. Sakuma, A. Nakano-Doi, T. Matsuyama, T. Nakagomi, Novel regenerative therapies based on regionally induced multipotent stem cells in post-stroke brains: their origin, characterization, and perspective, *Transl. Stroke Res.* 8 (6) (2017) 515–528.
- [149] K. Tatebayashi, Y. Tanaka, A. Nakano-Doi, R. Sakuma, S. Kamachi, M. Shirakawa, K. Uchida, H. Kageyama, T. Takagi, S. Yoshimura, T. Matsuyama, T. Nakagomi, Identification of multipotent stem cells in human brain tissue following stroke, *Stem Cells Dev.* 26 (11) (2017) 787–797.
- [150] C.S. Bjornsson, M. Apostolopoulou, Y. Tian, S. Temple, It takes a village: constructing the neurogenic niche, *Dev. Cell* 32 (4) (2015) 435–446.
- [151] C. Zhao, W. Deng, F.H. Gage, Mechanisms and functional implications of adult neurogenesis, *Cell* 132 (4) (2008) 645–660.
- [152] J. Song, C. Zhong, M.A. Bonaguidi, G.J. Sun, D. Hsu, Y. Gu, K. Meletis, Z.J. Huang, S. Ge, G. Enikolopov, K. Deisseroth, B. Luscher, K.M. Christian, G.L. Ming, H. Song, Neuronal circuitry mechanism regulating adult quiescent neural stem-cell fate decision, *Nature* 489 (7414) (2012) 150–154.
- [153] J. Song, K.M. Christian, G.L. Ming, H. Song, Modification of hippocampal circuitry by adult neurogenesis, *Dev. Neurobiol.* 72 (7) (2012) 1032–1043.
- [154] J.P. Andreotti, P.H.D.M. Prazeres, L.A.V. Magno, M.A. Romano-Silva, A. Mintz, A. Birbrair, Neurogenesis in the postnatal cerebellum after injury, *Int. J. Dev. Neurosci.* (2018).
- [155] J. Song, J. Sun, J. Moss, Z. Wen, G.J. Sun, D. Hsu, C. Zhong, H. Davoudi, K.M. Christian, N. Toni, G.L. Ming, H. Song, Parvalbumin interneurons mediate neuronal circuitry-neurogenesis coupling in the adult hippocampus, *Nat. Neurosci.* 16 (12) (2013) 1728–1730.
- [156] J. Song, R.H. Olsen, J. Sun, G.L. Ming, H. Song, Neuronal circuitry mechanisms regulating adult mammalian neurogenesis, *Cold Spring Harb. Perspect. Biol.* 8 (8) (2016).
- [157] O.W. Howell, H.E. Scharfman, H. Herzog, L.E. Sundstrom, A. Beck-Sickingler, W.P. Gray, Neuropeptide Y is neuroproliferative for post-natal hippocampal precursor cells, *J. Neurochem.* 86 (3) (2003) 646–659.
- [158] O.W. Howell, S. Silva, H.E. Scharfman, A.A. Sosunov, M. Zaben, A. Shtaya, G. McKhann 2nd, H. Herzog, A. Laskowski, W.P. Gray, Neuropeptide Y is important for basal and seizure-induced precursor cell proliferation in the hippocampus, *Neurobiol. Dis.* 26 (1) (2007) 174–188.
- [159] A. Cardoso, P. Freitas-da-Costa, L.S. Carvalho, N.V. Lukoyanov, Seizure-induced changes in neuropeptide Y-containing cortical neurons: potential role for seizure threshold and epileptogenesis, *Epilepsy Behav.: E&B* 19 (4) (2010) 559–567.
- [160] M. Zaben, W.J. Sheward, A. Shtaya, C. Abbosh, A.J. Harmar, A.K. Pringle, W.P. Gray, The neurotransmitter VIP expands the pool of symmetrically dividing postnatal dentate gyrus precursors via VPAC2 receptors or directs them toward a neuronal fate via VPAC1 receptors, *Stem Cells* 27 (10) (2009) 2539–2551.
- [161] H. Bao, B. Asrican, W. Li, B. Gu, Z. Wen, S.A. Lim, I. Haniff, C. Ramakrishnan, K. Deisseroth, B. Philpot, J. Song, Long-range GABAergic inputs regulate neural stem cell quiescence and control adult hippocampal neurogenesis, *Cell Stem Cell* 21 (5) (2017) 604–617 e5.
- [162] C. Romero-Grimaldi, B. Moreno-Lopez, C. Estrada, Age-dependent effect of nitric oxide on subventricular zone and olfactory bulb neural precursor proliferation, *J. Comp. Neurol.* 506 (2) (2008) 339–346.
- [163] C.K. Tong, J. Chen, A. Cebrian-Silla, Z. Mirzadeh, K. Obernier, C.D. Guinto, L.H. Tecott, J.M. Garcia-Verdugo, A. Kriegstein, A. Alvarez-Buylla, Axonal control of the adult neural stem cell niche, *Cell Stem Cell* 14 (4) (2014) 500–511.
- [164] P. Paez-Gonzalez, B. Asrican, E. Rodriguez, C.T. Kuo, Identification of distinct CHAT(+) neurons and activity-dependent control of postnatal SVZ neurogenesis, *Nat. Neurosci.* 17 (7) (2014) 934–942.
- [165] C.L. Lao, C.S. Lu, J.C. Chen, Dopamine D3 receptor activation promotes neural stem/progenitor cell proliferation through AKT and ERK1/2 pathways and expands type-B and -C cells in adult subventricular zone, *Glia* 61 (4) (2013) 475–489.
- [166] Y. Kim, W.Z. Wang, I. Comte, E. Pastrana, P.B. Tran, J. Brown, R.J. Miller, F. Doetsch, Z. Molnar, F.G. Szele, Dopamine stimulation of postnatal murine subventricular zone neurogenesis via the D3 receptor, *J. Neurochem.* 114 (3) (2010) 750–760.
- [167] B. Winner, P. Desplats, C. Hagl, J. Klucken, R. Aigner, S. Ploetz, J. Laemke, A. Karl, L. Aigner, E. Masliah, E. Buerger, J. Winkler, Dopamine receptor activation promotes adult neurogenesis in an acute Parkinson model, *Exp. Neurol.* 219 (2) (2009) 543–552.
- [168] D.A. Berg, L. Belnoue, H. Song, A. Simon, Neurotransmitter-mediated control of neurogenesis in the adult vertebrate brain, *Development* 140 (12) (2013) 2548–2561.
- [169] A. Perez-Villalba, M.S. Sirerol-Piquer, G. Belenguer, R. Soriano-Canton, A.B. Munoz-Manchado, J. Villadiego, D. Alarcon-Aris, F.N. Soria, B. Dehay, E. Bezar, M. Vila, A. Bortolozzi, J.J. Toledo-Aral, F. Perez-Sanchez, I. Farinas, Synaptic regulator alpha-synuclein in dopaminergic fibers is essentially required for the maintenance of subependymal neural stem cells, *J. Neurosci.* 38 (4) (2018) 814–825.
- [170] S. Ltic, M. Perovic, A. Mladenovic, N. Raicevic, S. Ruzdijic, L. Rakic, S. Kanazir, Alpha-synuclein is expressed in different tissues during human fetal development, *J. Mol. Neurosci.* 22 (3) (2004) 199–204.
- [171] A. Paul, Z. Chaker, F. Doetsch, Hypothalamic regulation of regionally distinct adult neural stem cells and neurogenesis, *Science* 356 (6345) (2017) 1383–1386.
- [172] J.P. Andreotti, L. Lousado, L.A.V. Magno, A. Birbrair, Hypothalamic neurons take center stage in the neural stem cell niche, *Cell Stem Cell* 21 (3) (2017) 293–294.
- [173] R.G. Allen, B. Peng, M.J. Pellegrino, E.D. Miller, D.K. Grandy, J.R. Lundblad, C.L. Washburn, J.E. Pintar, Altered processing of pro-orphanin FQ/nociceptin and pro-opiomelanocortin-derived peptides in the brains of mice expressing defective prohormone convertase 2, *J. Neurosci.* 21 (16) (2001) 5864–5870.
- [174] B.W. Lindsey, V. Tropepe, A comparative framework for understanding the biological principles of adult neurogenesis, *Prog. Neurobiol.* 80 (6) (2006) 281–307.
- [175] O. Bergmann, K.L. Spalding, J. Frisen, Adult neurogenesis in humans, *Cold Spring Harb. Perspect. Biol.* 7 (7) (2015) a018994.
- [176] A. Quinones-Hinojosa, N. Sanai, M. Soriano-Navarro, O. Gonzalez-Perez, Z. Mirzadeh, S. Gil-Perotin, R. Romero-Rodriguez, M.S. Berger, J.M. Garcia-Verdugo, A. Alvarez-Buylla, Cellular composition and cytoarchitecture of the adult human subventricular zone: a niche of neural stem cells, *J. Comp. Neurol.* 494 (3) (2006) 415–434.
- [177] A. Ernst, K. Alkass, S. Bernard, M. Salehpour, S. Perl, J. Tisdale, G. Possnert, H. Druid, J. Frisen, Neurogenesis in the striatum of the adult human brain, *Cell* 156 (5) (2014) 1072–1083.
- [178] O. Bergmann, J. Liebl, S. Bernard, K. Alkass, M.S. Yeung, P. Steier, W. Kutschera, L. Johnson, M. Landen, H. Druid, K.L. Spalding, J. Frisen, The age of olfactory bulb neurons in humans, *Neuron* 74 (4) (2012) 634–639.
- [179] M. Batailler, M. Drogue, M. Baroncini, C. Fontaine, V. Prevot, M. Migaud, DCX-expressing cells in the vicinity of the hypothalamic neurogenic niche: a comparative study between mouse, sheep, and human tissues, *J. Comp. Neurol.* 522 (8) (2014) 1966–1985.
- [180] A.B. Nogueira, M.C. Sogayar, A. Colquhoun, S.A. Siqueira, A.B. Nogueira, P.E. Marchiori, M.J. Teixeira, Existence of a potential neurogenic system in the adult human brain, *J. Transl. Med.* 12 (2014) 75.
- [181] K.L. Spalding, O. Bergmann, K. Alkass, S. Bernard, M. Salehpour, H.B. Huttner, E. Bostrom, I. Westerlund, C. Vial, B.A. Buchholz, G. Possnert, D.C. Mash, H. Druid, J. Frisen, Dynamics of hippocampal neurogenesis in adult humans, *Cell* 153 (6) (2013) 1219–1227.
- [182] R. Knott, I. Singec, M. Ditter, F. Pantazis, P. Capetian, R.P. Meyer, V. Horvat, B. Volk, G. Kempermann, Murine features of neurogenesis in the human hippocampus across the lifespan from 0 to 100 years, *PLoS One* 5 (1) (2010) e8809.
- [183] S.F. Sorrells, M.F. Paredes, A. Cebrian-Silla, K. Sandoval, D. Qi, K.W. Kelley, D. James, S. Mayer, J. Chang, K.I. Auguste, E.F. Chang, A.J. Gutierrez, A.R. Kriegstein, G.W. Mathern, M.C. Oldham, E.J. Huang, J.M. Garcia-Verdugo, Z. Yang, A. Alvarez-Buylla, Human hippocampal neurogenesis drops sharply in children to undetectable levels in adults, *Nature* 555 (7696) (2018) 377–381.
- [184] M. Boldrini, C.A. Fulmore, A.N. Tartt, L.R. Simeon, I. Pavlova, V. Poposka, G.B. Rosoklija, A. Stankov, V. Arango, A.J. Dwork, R. Hen, J.J. Mann, Human hippocampal neurogenesis persists throughout aging, *Cell Stem Cell* 22 (4) (2018) 589–599 e5.
- [185] G. Kempermann, F.H. Gage, L. Aigner, H. Song, M.A. Curtis, S. Thuret, H.G. Kuhn, S. Jessberger, P.W. Frankland, H.A. Cameron, E. Gould, R. Hen, D.N. Abrous, N. Toni, A.F. Schinder, X. Zhao, P.J. Lucassen, J. Frisen, Human adult neurogenesis: evidence and remaining questions, *Cell Stem Cell* 23 (1) (2018) 25–30.
- [186] F. Henriques, et al., Toll-like receptor-4 disruption suppresses adipose tissue remodeling and increases survival in cancer cachexia syndrome, *Sci. Rep.* 8 (2018) 18024, <https://doi.org/10.1038/s41598-018-36626-3>.
- [187] L.X. Pereira, et al., Synthetic matrix of polyether-polyurethane as a biological platform for pancreatic regeneration, *Life Sci.* 176 (2017) 67–74, <https://doi.org/10.1016/j.lfs.2017.03.015>.
- [188] A. Birbrair, Stem cells heterogeneity, *Adv. Exp. Med. Biol.* (2019).
- [189] A.B. Salmina, Y.K. Komleva, O.L. Lopatina, A. Birbrair, Pericytes in Alzheimer's Disease: novel clues to cerebral amyloid angiopathy pathogenesis, *Adv. Exp. Med. Biol.* (2019).
- [190] R.S.N. Barreto, et al., Pericytes in the placenta: role in placental development and homeostasis, *Adv. Exp. Med. Biol.* (2019).