ELSEVIER

Contents lists available at ScienceDirect

Experimental Gerontology

journal homepage: www.elsevier.com/locate/expgero



Review

How increased oxidative stress promotes longevity and metabolic health: The concept of mitochondrial hormesis (mitohormesis)

Michael Ristow a,b,*, Kim Zarse a

- ^a Dept. of Human Nutrition, Institute of Nutrition, University of Jena, 29 Dornburger Str., Jena D-07743, Germany
- ^b Dept. of Clinical Nutrition, German Institute of Human Nutrition, 114 Arthur-Scheunert-Allee, Nuthetal D-14558, Germany

ARTICLE INFO

Article history: Received 1 July 2009 Received in revised form 9 March 2010 Accepted 19 March 2010 Available online 27 March 2010

Keywords:
Aging
Metabolism
Reactive oxygen species
Oxidative stress
Glucose
Glycolysis
Calorie restriction

ABSTRACT

Recent evidence suggests that calorie restriction and specifically reduced glucose metabolism induces mitochondrial metabolism to extend life span in various model organisms, including Saccharomyces cerevisiae, Drosophila melanogaster, Caenorhabditis elegans and possibly mice. In conflict with Harman's free radical theory of aging (FRTA), these effects may be due to increased formation of reactive oxygen species (ROS) within the mitochondria causing an adaptive response that culminates in subsequently increased stress resistance assumed to ultimately cause a long-term reduction of oxidative stress. This type of retrograde response has been named mitochondrial hormesis or mitohormesis, and may in addition be applicable to the health-promoting effects of physical exercise in humans and, hypothetically, impaired insulin/IGF-1-signaling in model organisms. Consistently, abrogation of this mitochondrial ROS signal by antioxidants impairs the lifespan-extending and health-promoting capabilities of glucose restriction and physical exercise, respectively. In summary, the findings discussed in this review indicate that ROS are essential signaling molecules which are required to promote health and longevity. Hence, the concept of mitohormesis provides a common mechanistic denominator for the physiological effects of physical exercise, reduced calorie uptake, glucose restriction, and possibly beyond.

© 2010 Elsevier Inc. All rights reserved.

1. Calorie restriction

A limited reduction of nutritional calorie uptake, so-called calorie restriction (CR), has been shown to extend life span in multiple species and model organisms, as initially observed by McCay et al. (1935). It is beyond the scope of this review to summarize the multiple findings on CR, since excellent reviews on this topic have been published in the past (Weindruch and Walford, 1988; Masoro, 2000; Speakman et al., 2002; Heilbronn and Ravussin, 2003; Ingram et al., 2004; Anson et al., 2005; Gredilla and Barja, 2005; Sinclair, 2005; Wolff and Dillin, 2006; Bishop and Guarente, 2007; Piper and Bartke, 2008). It should be emphasized, however, that unequivocal evidence for the effectiveness of CR in primates and especially humans is missing. A recent publication on an ongoing study in Macaca mulatta shows that CR has no statistically significant effect on overall mortality (Colman et al., 2009). However, since about half of the study group was still alive at the time of manuscript preparation, future findings from this ongoing study may show whether CR in rhesus monkeys significantly affects mortality. Nevertheless, so-called "age-related mortality" was signifi-

E-mail address: mristow@mristow.org (M. Ristow).

cantly decreased in *M. mulatta*. It should be noted, though, that age-related mortality (as defined in this study) accounted for only 54% of deaths during the study period. In contrast and quite strikingly, age-related gluco-regulatory impairment was completely abolished in calorically restricted monkeys. Hence and due to additional findings (Fontana et al., 2004; Heilbronn et al., 2006; Ingram et al., 2006a; Weindruch, 2006; Fontana and Klein, 2007) it appears possible that CR extends life span in primates and/or humans.

The initial conceptual background for restricting dietary calories is based on the assumption that reducing nutritive calorie availability would reduce the metabolic rate of an organism. Accordingly, it was proposed more than a century ago that maximum life span is inversely proportional to the amount of nutritive energy metabolized (Rubner, 1908). Subsequently, the rate-of-living hypothesis evolved, suggesting that an increased metabolic rate would decrease life span in eukaryotes (Pearl, 1928). Several decades later it was proposed that increased metabolic rate would promote increased formation of reactive oxygen species (ROS) to cause cumulative damage to the cell, and hence the organism (Harman, 1956). Notably, respiratory enzymes using oxygen to generate readily available energy were explicitly proposed to be the most relevant culprit in this regard (Harman, 1956). This concept was named free radical theory of aging (FRTA).

Based on these assumptions, considerable experimental effort has been made to elucidate the underlying mechanistic principles.

^{*} Corresponding author at: Dept. of Human Nutrition, Institute of Nutrition, University of Jena, 29 Dornburger Str., Jena D-07743, Germany. Tel.: +49 3641 949630; fax: +49 3641 949632.

On the one hand, it has repeatedly been shown that CR is capable of delaying a number of age-related diseases, including obesity, type 2 diabetes, hypercholesterolemia, atherosclerosis, different cancers, as well as neurodegeneration and cardiomyopathy. This has been attributed to specific and diverse effects of CR on the respective molecular processes assumed to cause these disorders. According to some of these approaches, delayed aging would simply reflect a cumulative reduction of age-associated and mortalitypromoting medical conditions as a consequence of CR. On the other hand, it was shown that CR per se promotes increased stress defense, and specifically induces endogenous defense mechanisms against ROS (Koizumi et al., 1987; Semsei et al., 1989; Rao et al., 1990; Pieri et al., 1992; Youngman et al., 1992; Xia et al., 1995; Masoro, 1998a; Barros et al., 2004). In most cases, this was interpreted as a consequence of reduced metabolic rate, and hence reduced ROS production. More recently, a different perspective has emerged, suggesting that CR causes an adaptive response to specific metabolic alterations in states of reduced food uptake.

2. Reduction of specific macronutrients

Nutritional, *i.e.* metabolizable calories are derived from carbohydrates, triacylglycerols (fat) and proteins. These contain a few different monosaccharides (including glucose), and significant numbers of fatty acids and amino acids, respectively. Limited evidence exists whether the generally accepted effects of calorie restriction can be attributed to specific macronutrients, *i.e.* whether restriction of a single macronutrient may exert the same effects than overall CR does. This topic has been reviewed in detail elsewhere (Piper and Bartke, 2008), hence the following paragraphs will focus on specific aspects of macronutrient choice only.

In invertebrate model organisms, restriction of proteins as well as carbohydrates, mostly glucose, has been studied with different and sometimes opposing outcomes, whereas studies on triacylglycerols are lacking for invertebrates. While the effects of glucose restriction will be discussed below, in *Drosophila melanogaster*, restriction of casein extends life span (Min and Tatar, 2006), Moreover, it has been proposed that restriction of both yeast as well as sugar may extend Drosophila lifespan despite unaltered calorie uptake (Mair et al., 2005). Very recently, it was shown that increased abundance of essential amino acids, and particularly methionine counteracts the lifespan-extending effects of CR in D. melanogaster (Grandison et al., 2009). Notably, restriction of methionine in rodents similarly delays ageing (Zimmerman et al., 2003; Miller et al., 2005) and increasing protein content impairs antioxidant defense in rats (De et al., 1983). For Caenorhabditis elegans, impaired activity of peptide transport similarly extends life span (Meissner et al., 2004). However, selective depletion of nutritive amino acids is difficult to achieve in C. elegans, and hence to our best knowledge has not been studied.

In mammals and especially humans, increasing evidence suggests that a number of health-promoting metabolic effects can by more easily achieved by a selective reduction of dietary carbohydrates: Whereas efficacy of long-term weight reduction appears to be comparable between low-carbohydrate and low-energy diets (mainly depleted in triacylglycerols) (Nordmann et al., 2006; Hession et al., 2009), several serum parameters seem to be favourably affected by a specific reduction of carbohydrate uptake, whereas total energy uptake was, in most studies, not significantly affected by the type of diet (Nordmann et al., 2006; Hession et al., 2009). Hence, it appears feasible that a depletion of carbohydrates and/or glucose only exerts specific effects beyond those observed with general CR. In anticipation of mechanisms outlined below, it should be noted that metabolism of glucose can yield ATP even in the absence of mitochondrial organelles or even oxygen, while

conversion of fatty acids and/or (most) amino acids into ATP depends on oxidative phosphorylation (OxPhos) and hence oxygen.

3. Glucose restriction

A specific restriction of nutritive glucose is, with the exception of yeast and *D. melanogaster*, difficult to achieve in eukaryotic model organisms. In *Saccharomyces cerevisiae*, it was shown that reduced glucose availability significantly extends chronological life span, and this extension depends on induction of respiration (Lin et al., 2002) as well as sirtuins (Lin et al., 2000). While the dependence on sirtuins is a matter of debate (Kaeberlein et al., 2004; Agarwal et al., 2005; Guarente, 2006; Smith et al., 2007), alternative mechanisms independent of sirtuins have been proposed (Barros et al., 2004; Roux et al., 2009).

In C. elegans and mammals, a specific restriction of intracellular glucose availability is commonly achieved by application of a competitive inhibitor of glycolysis, 2-deoxy-glucose (DOG) (Wick et al., 1957). In C. elegans, it was shown that application of DOG induces respiration and extends life span (Schulz et al., 2007), in this regard reflecting previous findings in yeast (Lin et al., 2002). However and in conflict with these aforementioned findings in yeast, this process was independent of sirtuins, but rather required activation of AMP-activated kinase (AMPK) (Schulz et al., 2007). This kinase was previously established as a sensor of cellular energy depletion in both mammals (Hardie et al., 2006) and specifically C. elegans (Apfeld et al., 2004; Greer et al., 2007), and has been found to induce a health-promoting metabolic state particularly by inducing mitochondrial metabolism (Hardie et al., 2006). Accordingly, application of DOG to rodents efficiently mimics features of the metabolic state of CR (Lane, 1998) as well as carbohydrate restriction (Garriga-Canut et al., 2006), suggesting that DOG acts as a CR mimetic (Duan and Mattson, 1999; Sinclair, 2005; Zhu et al., 2005; Ingram et al., 2006b).

Accordingly and as an alternate route to modulate intracellular glucose availability, combined disruption of insulin-dependent glucose transporter GLUT4 in adipose and muscle tissues of mice causes adult hyperglycemia as well as a metabolic switch to increased fatty acid turnover and utilization, while lifespan was studied up to 18 months of age only, and found to be unaltered (Kotani et al., 2004). Conversely, transgenic over-expression of GLUT4 in mice efficiently lowers blood glucose by increasing cellular glucose uptake, but does not extend life span (McCarter et al., 2007). Moreover, it was shown that increased glucose availability reduces C. elegans lifespan (Schulz et al., 2007) while potentially underlying mechanisms have been subsequently proposed (Lee et al., 2009; Schlotterer et al., 2009). Altogether, these findings suggest that increased intracellular glucose availability exerts detrimental effects on longevity, whereas decreased glucose availability promotes oxidative metabolism and extends life span.

4. Impaired insulin/IGF-1 signaling and glucose availability

Insulin and insulin-like growth factor 1 (IGF-1) are peptide hormones. Insulin is produced in and secreted from the pancreatic beta-cells, while IGF-1 is produced in the liver. IGF-1 production and release depends on a third hormone named somatotropin (STH) a.k.a. growth hormone (GH) which stems from the anterior pituitary gland. Insulin, GH and IGF-1 are hormones that bind to specific and, at least in mammals, distinct receptors. However, GH exerts some of its effects indirectly by regulating the abundance of IGF-1. Moreover, it should be noted that most of the IGF-1-independent, *i.e.* direct and receptor-mediated effects of GH commonly counteract insulin action.

Impaired availability and/or activity of GH and/or IGF-1 starting in early life causes reduced growth or dwarfism. Mice with the corresponding mutations are called Ames, Snell, and *little*, and have been described in more detail elsewhere (Quarrie and Riabowol, 2004). Interestingly, such growth-impaired mice have increased lifespan (Brown-Borg et al., 1996), while increased GH signaling impairs lifespan (Pendergrass et al., 1993; Steger et al., 1993).

Accordingly, heterozygous global disruption of the IFG-1 receptor (Holzenberger et al., 2003) as well as impaired neuronal IFG-1 receptor function (Kappeler et al., 2008) extend murine lifespan, and prevents proteotoxicity and neurodegeneration (Cohen et al., 2009).

Impaired activation of the insulin receptor has been linked to a state called insulin resistance, defined as an inappropriately reduced intracellular response to an extracellular insulin stimulus (Kahn, 1994). The key intracellular response towards extracellular activation of the insulin receptor is increased glucose uptake as mediated by translocation of the glucose transporter GLUT4. Hence it appears generally accepted that insulin resistance causes type 2 diabetes leading to reduced intracellular glucose availability (Biddinger and Kahn, 2006). This notion is supported by observations from humans in regards to increased prevalence of hyperglycemia, insulin resistance and lastly type 2 diabetes with increasing age (DeFronzo, 1981).

Conversely, targeted whole-body disruption of the insulin receptor in mice causes embryonic lethality, but when disruption is restricted to the (in regards to glucose metabolism) most relevant tissue, skeletal muscle, neither hyperglycemia nor diabetes was observed, but rather a striking increase in fatty acid turnover occurred (Brüning et al., 1998). Life span analyses in these mice have not been published. In addition, adipocyte-specific disruption of the insulin receptor extends murine life span (Blüher et al., 2003) and so does global heterozygous disruption of the downstream insulin receptor substrate 1 (IRS-1) (Selman et al., 2008a) which interestingly also is located downstream of the of IGF-1 receptor. Similarly, neuronal disruption of IRS-2 was shown to promote longevity (Taguchi et al., 2007), and so did heterozygous global disruption of IRS-2 (Taguchi et al., 2007) while others could not confirm the latter evidence using the same model (Selman et al., 2008b). Taken together, these findings suggest that a limited impairment of insulin and/or IGF-1 signaling may actually extend murine life span due to widely unresolved reasons. Of note, mutations of insulin/IGF-1 signaling have been shown to be associated with human longevity (van Heemst et al., 2005; Pawlikowska et al., 2009).

In invertebrates, long-standing evidence exists in this regard: In both *C. elegans* and *D. melanogaster*, mutations in the respective orthologues of the insulin/IGF-1 receptor or proteins located downstream of these receptors significantly extend life span (Friedman and Johnson, 1988; Kenyon et al., 1993; Kimura et al., 1997; Clancy et al., 2001; Tatar et al., 2001). However, there is little evidence in invertebrates whether and to which extent impaired insulin/IGF-1 signaling affects glucose availability.

While some authors propose that impaired insulin/IGF-1/GH signaling extends life span independently of pathways activated by CR (Lakowski and Hekimi, 1998; Bartke et al., 2001; Houthoofd et al., 2003; Min et al., 2008; Bonkowski et al., 2009), others have suggested that impaired insulin/IGF-1 signaling may share mechanistic features of caloric restriction and hence decreased energy availability, at least to some extent (Brown-Borg et al., 2002; Clancy et al., 2002; Al-Regaiey et al., 2005; Bonkowski et al., 2006; Greer et al., 2007; Narasimhan et al., 2009; Yen and Mobbs, in press). Independently, it appears likely that impaired insulin/IGF-1 signaling causes an intracellular glucose depletion in most model organisms, hypothetically mimicking the metabolic state of glucose restriction, hence contributing to lifespan extension by impaired insulin/IGF-1 signaling. While experimental evidence for this hypothesis is missing, some findings from rodents support

the assumption that impaired insulin/IGF-1 signaling induces mitochondrial metabolism, whereas lifespan in most cases has not been studied (Yechoor et al., 2004; Brooks et al., 2007; Katic et al., 2007; Russell and Kahn, 2007; Westbrook et al., 2009).

5. Induction of mitochondrial metabolism by calorie/glucose restriction

While some papers suggest that the net uptake of calories is not reduced over life time in states of CR (Masoro et al., 1982; Mair et al., 2005), it is by definition agreed upon that during the actual CR intervention a relative depletion of available energy occurs.

Mitochondria convert nutritional energy more effectively into readily available energy, i.e. ATP, than non-oxidative metabolism of carbohydrates and some amino acids does. E.g., while glycolytic metabolism of one mol of glucose generates 4 mols of ATP only, its oxidative metabolism generates 30 mols of ATP. Hence, and as indicated by findings in yeast (Lin et al., 2002) and C. elegans (Schulz et al., 2007), decreased glucose availability would induce mitochondrial metabolism to increase OxPhos, aiming to maintain intracellular ATP supply. Similarly, however analyzing global CR (and not specifically glucose restriction), it was shown that food deprivation promotes mitochondrial biogenesis and OxPhos in rodents (Nisoli et al., 2005). Additionally, it has been suggested that mass-specific energy expenditure in CR rats is higher than expected (Selman et al., 2005) and that cultured mammalian cells induce their respiratory capacity in states of CR (Lopez-Lluch et al., 2006). Moreover, impaired insulin/IGF-1/GH signaling causes an induction of mitochondrial metabolism in rodents (Yechoor et al., 2004; Katic et al., 2007; Russell and Kahn, 2007; Westbrook et al., 2009). In addition, induction of mitochondrial metabolism by various pharmacological measures (Ames, 2005) and specifically physical exercise (Warburton et al., 2006; Lanza et al., 2008) has been proposed to extend lifespan. In contrast, mitochondrial dysfunction has been proposed as a key cause of aging (Trifunovic and Larsson, 2008; Bratic and Trifunovic, 2010), diabetes (Wiederkehr and Wollheim, 2006), cancer (Ristow, 2006), as well as neurodegeneration (Fukui and Moraes, 2008; Tatsuta and Langer, 2008). Moreover, impaired mitochondrial capacity decreases life span in yeast (Bonawitz et al., 2006), C. elegans (Zarse et al., 2007) and rodents (Thierbach et al., 2005). Mechanistically, sirtuins (see above) as well as AMPK signaling (see above) may be involved. Moreover, disruption of the targetof-rapamycin protein mTOR (Wullschleger et al., 2006) has been shown to extend S. cerevisae lifespan (Powers et al., 2006) interestingly by inducing mitochondrial metabolism (Bonawitz et al., 2007). Consistently and as to be anticipated from states of glucose restriction (Schulz et al., 2007) as well as TOR disruption (Bonawitz et al., 2007), the TOR target and translational repressor 4E-BP has subsequently been shown to modulate mitochondria metabolism in states of CR (Zid et al., 2009). Moreover, TOR/TORC1 activity appears to be controlled by AMPK (Gwinn et al., 2008), altogether suggesting that both TOR and AMPK may be upstream regulators of mitochondrial metabolism and OxPhos.

While all these aforementioned findings suggest that increased mitochondrial metabolism is instrumental and possibly required for the extension of lifespan, it should be noted that, in conflict with the findings mentioned above, CR has been shown to increase life span in the absence of increased respiration (Houthoofd et al., 2002; Kaeberlein et al., 2004), or even in the absence of respiration at all (Kaeberlein et al., 2005).

6. Oxidative stress and mitochondrial hormesis (mitohormesis)

About five decades ago and as stated above, increased formation of ROS as a consequence of increased metabolic rate was pro-

posed to be the major culprit for the ageing process and decreased life span (Harman, 1956). Mitochondria are the main source of ROS. For a long time, these were considered exclusively unwanted by-products of OxPhos. In support of this view, a significant number of studies in various model organisms suggests that amelioration of oxidative stress contributes to an increase of lifespan (Harrington and Harley, 1988; Phillips et al., 1989; Orr and Sohal, 1994; Parkes et al., 1998; Melov et al., 1999; Adachi and Ishii, 2000; Melov et al., 2000; Moskovitz et al., 2001; Bakaev and Lyudmila, 2002; Ruan et al., 2002; Ishii et al., 2004; Huang et al., 2006; Zou et al., 2007; Kim et al., 2008; Quick et al., 2008; Dai et al., 2009; Shibamura et al., 2009). Consistently, significant effort has been made to reduce ROS formation due to the assumption that such interventions may block or at least ameliorate aging processes in humans.

Accordingly, both synthetic as well as naturally occurring compounds that physically interact with ROS to inactivate the latter. so-called antioxidants, have been extensively been investigated. Unexpectedly, many prospective clinical trials aiming to find any health-promoting effects of antioxidants failed, in the best case showing no health-promoting effects of these compounds (Greenberg et al., 1994; Liu et al., 1999; Rautalahti et al., 1999; Virtamo et al., 2000; Heart Protection Study Collaborative Group, 2002; Sacco et al., 2003; Zureik et al., 2004; Czernichow et al., 2005, 2006; Cook et al., 2007; Kataja-Tuomola et al., 2008; Sesso et al., 2008; Katsiki and Manes, 2009; Lin et al., 2009; Song et al., 2009). More importantly, a number of studies suggests that antioxidants may promote cancer in humans (Bjelakovic et al., 2004; Bairati et al., 2005; Hercberg et al., 2007; Bardia et al., 2008; Lawenda et al., 2008; Myung et al., 2010). Accordingly, other studies show that antioxidant supplements may be disease-promoting and/or may even reduce lifespan in humans (Albanes et al., 1996; Omenn et al., 1996; Vivekananthan et al., 2003; Lonn et al., 2005; Bjelakovic et al., 2007; Ward et al., 2007; Lippman et al., 2009).

Consistently and in conflict with Harman's hypothesis, evidence has emerged in recent years that ROS may actually work as essential, and potentially lifespan-promoting, signaling molecules which transduce signals from the mitochondrial compartment to other compartments of the cell (Barja, 1993; Rhee et al., 2003; Kaelin, 2005; Connor et al., 2005; Guzy et al., 2005; Guzy and Schumacker, 2006; Chandel and Budinger, 2007; Schulz et al., 2007; Veal et al., 2007; Owusu-Ansah et al., 2008; Finley and Haigis, 2009; Ristow et al., 2009; Loh et al., 2009). Independently, it has been suggested that CR acts by inducing low-level stress that culminates in increased stress resistance and ultimately longevity (Masoro, 1998b,a). This would reflect an adaptive response commonly defined as hormesis (Southam and Ehrlich, 1943) (for a current definition see (Calabrese et al., 2007)), and was later named mitochondrial hormesis or mitohormesis, referring to ROS-related stress emanating from the mitochondria (Tapia, 2006).

Consistent with these hypotheses, it was shown in rodents that calorie restriction induces antioxidant defense capacities (Koizumi et al., 1987; Semsei et al., 1989; Rao et al., 1990; Pieri et al., 1992; Youngman et al., 1992; Xia et al., 1995; Sreekumar et al., 2002). In yeast, glucose restriction decreases ROS production despite in increased respiratory activity (Barros et al., 2004). In contrast and while using the same model organism, others showed that glucose restriction increases ROS production (Agarwal et al., 2005; Kharade et al., 2005; Piper et al., 2006). Interestingly, an induction of ROS defense enzymes was also observed (Agarwal et al., 2005; Kharade et al., 2005; Piper et al., 2006), tentatively suggesting a mechanistic link between increased respiration, elevated ROS production and adaptive induction of ROS defense.

Accordingly, in *D. melanogaster* CR was unable to primarily decrease ROS production, and genetically decreased ROS production was unable to extend life span (Miwa et al., 2004). Consistently,

altering ROS production or antioxidant defense in various model organisms has similarly failed to reciprocally modulate life span (Huang et al., 2000; Bayne and Sohal, 2002; Keaney and Gems, 2003; Andziak et al., 2006; Selman et al., 2006; Ran et al., 2007; Doonan et al., 2008; Heidler et al., 2009; Jang and van Remmen, 2009; Jang et al., 2009; Lapointe et al., 2009; Van Raamsdonk and Hekimi, 2009; Yen et al., 2009; Zhang et al., 2009; Pun et al., 2010). Moreover, long-lived mutants of C. elegans unambiguously show increased stress resistance which in some studies is paralleled by increased metabolic activity (Lithgow et al., 1995; Vanfleteren and De Vreese, 1995; Honda and Honda, 1999; Murphy et al., 2003; Houthoofd et al., 2005; Dong et al., 2007). Of note, similar results in regards to neuroprotective mechanisms of CR and specifically DOG application have been described in rodents (Arumugam et al., 2006). Lastly, humans on a ketogenic, i.e. carbohydrate-depleted diet show increased antioxidant defense presumably following increased oxidative metabolism due to increased rates of beta-oxidation (Nazarewicz et al., 2007).

While all these aforementioned publications support the possibility that ROS itself induce ROS defense and ultimately increase life span, it remained to be shown that prevention of ROS formation would reduce the life-extending capabilities of CR. These experiments were undertaken by showing that DOG reduces glucose availability, increases respiration and ROS formation, promotes activity of ROS defense enzymes, and extends life span in *C. elegans* (Schulz et al., 2007). Notably, co-treatment of nematodes with several different antioxidants which inactivate ROS fully abolished the life-extending effects of CR and DOG, providing direct evidence for an essential role of increased ROS formation in extension of life span (Schulz et al., 2007).

7. Physical exercise

As summarized above, CR and specifically glucose restriction induce mitochondrial respiration and ROS formation in various model organisms. The ROS signal appears to induce ROS defense mechanisms, culminating in extended lifespan, which reflects a typical adaptive response, consistent with the mitohormesis hypothesis. Antioxidants prevent this adaptive response, and extension of lifespan is abolished. It remains to be resolved, in which time-resolved order these processes occur, and specifically whether increased ROS defense counteracts respiration-derived ROS formation.

Moreover, these findings indicate that approaches to induce mitochondrial metabolism are likely to promote metabolic health and may potentially extended lifespan. This notion is supported by the fact that not only calorie and/or glucose (and possibly amino acid) restriction, but also longevity-promoting physical exercise induces mitochondrial metabolism and ROS formation (Davies et al., 1982; Chevion et al., 2003; Powers and Jackson, 2008). Notably, supplementation with ROS-reducing antioxidants inhibits (Gomez-Cabrera et al., 2008; Ristow et al., 2009) the health-promoting effects (Higuchi et al., 1985; Lindsted et al., 1991; Manini et al., 2006; Warburton et al., 2006; Lanza et al., 2008) of physical exercise. This suggests that CR, glucose restriction and physical exercise share, at least in part, a common metabolic denominator (Fig. 1), i.e. increased mitochondrial metabolism and ROS formation inducing a adaptive response that culminates in increased stress resistance, antioxidant defense and extended life span.

Acknowledgments

Studies in the authors' laboratory have been or are supported by the German Research Association (DFG), the German Ministry of Education and Research (BMBF), the European Foundation for the

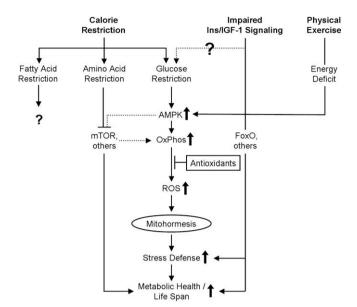


Fig. 1. Mitohormesis and lifespan extension: For both calorie restriction as well as physical exercise, experimental evidence suggests that induction of mitochondrial metabolism is required for the lifespan-extending and/or health-promoting effects of these interventions. This increase in mitochondrial metabolism generates a ROS signal that is required to induce an adaptive response to culminate in increased lifespan. For impaired insulin-IGF-1 signaling however, this links remains to be experimentally shown.

Study of Diabetes (EFSD), the Leibniz Association (WGL), the Fritz-Thyssen-Stiftung and the Wilhelm-Sander-Stiftung. We apologize to those whose work relevant for the topic has not been cited solely due to limitations of space.

References

- Adachi, H., Ishii, N., 2000. Effects of tocotrienols on life span and protein carbonylation in *Caenorhabditis elegans*. J. Gerontol. A Biol. Sci. Med. Sci. 55, B280–B285.
- Agarwal, S., Sharma, S., Agrawal, V., Roy, N., 2005. Caloric restriction augments ROS defense in S. cerevisiae by a Sir2p independent mechanism. Free Radic. Res. 39, 55–62
- Albanes, D., Heinonen, O.P., Taylor, P.R., Virtamo, J., Edwards, B.K., Rautalahti, M., Hartman, A.M., Palmgren, J., Freedman, L.S., Haapakoski, J., Barrett, M.J., Pietinen, P., Malila, N., Tala, E., Liippo, K., Salomaa, E.R., Tangrea, J.A., Teppo, L., Askin, F.B., Taskinen, E., Erozan, Y., Greenwald, P., Huttunen, J.K., 1996. Alpha-Tocopherol and beta-carotene supplements and lung cancer incidence in the alpha-tocopherol, beta-carotene cancer prevention study: effects of base-line characteristics and study compliance. J. Natl. Cancer. Inst. 88, 1560–1570.
- Al-Regaiey, K.A., Masternak, M.M., Bonkowski, M., Sun, L., Bartke, A., 2005. Long-lived growth hormone receptor knockout mice. Interaction of reduced insulin-like growth factor i/insulin signaling and caloric restriction. Endocrinology 146, 851–860.
- Ames, B.N., 2005. Increasing longevity by tuning up metabolism. To maximize human health and lifespan, scientists must abandon outdated models of micronutrients. EMBO Rep. 6 Spec No., S20–S24.
- Andziak, B., O'Connor, T.P., Qi, W., DeWaal, E.M., Pierce, A., Chaudhuri, A.R., Van Remmen, H., Buffenstein, R., 2006. High oxidative damage levels in the longest-living rodent, the naked mole-rat. Aging Cell 5, 463–471.
- Anson, R.M., Jones, B., de Cabo, R., 2005. The diet restriction paradigm: a brief review of the effects of every-other-day feeding. Age (Ohama) 27, 17–25.
- Apfeld, J., O'Connor, G., McDonagh, T., DiStefano, P.S., Curtis, R., 2004. The AMP-activated protein kinase aak-2 links energy levels and insulin-like signals to lifespan in C. elegans. Genes Dev. 18, 3004–3009.
- Arumugam, T.V., Gleichmann, M., Tang, S.C., Mattson, M.P., 2006. Hormesis/preconditioning mechanisms, the nervous system and aging. Ageing Res. Rev. 5, 165–178.
- Bairati, I., Meyer, F., Gelinas, M., Fortin, A., Nabid, A., Brochet, F., Mercier, J.P., Tetu, B., Harel, F., Masse, B., Vigneault, E., Vass, S., del Vecchio, P., Roy, J., 2005. A randomized trial of antioxidant vitamins to prevent second primary cancers in head and neck cancer patients. J. Natl. Cancer. Inst. 97, 481–488.
- Bakaev, V.V., Lyudmila, M.B., 2002. Effect of ascorbic acid on longevity in the nematoda Caenorhabditis elegans. Biogerontology 3 (Suppl. 1), 12–16.
- Bardia, A., Tleyjeh, I.M., Cerhan, J.R., Sood, A.K., Limburg, P.J., Erwin, P.J., Montori, V.M., 2008. Efficacy of antioxidant supplementation in reducing primary cancer

- incidence and mortality: systematic review and meta-analysis. Mayo Clin. Proc. 83, 23–34.
- Barja, G., 1993. Oxygen radicals, a failure or a success of evolution? Free Radic. Res. Commun. 18, 63–70.
- Barros, M.H., Bandy, B., Tahara, E.B., Kowaltowski, A.J., 2004. Higher respiratory activity decreases mitochondrial reactive oxygen release and increases life span in *Saccharomyces cerevisiae*. J. Biol. Chem. 279, 49883–49888.
- Bartke, A., Wright, J.C., Mattison, J.A., Ingram, D.K., Miller, R.A., Roth, G.S., 2001. Extending the lifespan of long-lived mice. Nature 414, 412.
- Bayne, A.C., Sohal, R.S., 2002. Effects of superoxide dismutase/catalase mimetics on life span and oxidative stress resistance in the housefly, *Musca domestica*. Free Radic. Biol. Med. 32, 1229–1234.
- Biddinger, S.B., Kahn, C.R., 2006. From mice to men: insights into the insulin resistance syndromes. Annu. Rev. Physiol. 68, 123–158.
- Bishop, N.A., Guarente, L., 2007. Genetic links between diet and lifespan: shared mechanisms from yeast to humans. Nat. Rev. Genet. 8, 835–844.
- Bjelakovic, G., Nikolova, D., Simonetti, R.G., Gluud, C., 2004. Antioxidant supplements for prevention of gastrointestinal cancers: a systematic review and meta-analysis. Lancet 364, 1219–1228.
- Bjelakovic, G., Nikolova, D., Gluud, L.L., Simonetti, R.G., Gluud, C., 2007. Mortality in randomized trials of antioxidant supplements for primary and secondary prevention: systematic review and meta-analysis. JAMA 297, 842–857.
- Blüher, M., Kahn, B.B., Kahn, C.R., 2003. Extended longevity in mice lacking the insulin receptor in adipose tissue. Science 299, 572–574.
- Bonawitz, N.D., Rodeheffer, M.S., Shadel, G.S., 2006. Defective mitochondrial gene expression results in reactive oxygen species-mediated inhibition of respiration and reduction of yeast life span. Mol. Cell. Biol. 26, 4818–4829.
- Bonawitz, N.D., Chatenay-Lapointe, M., Pan, Y., Shadel, G.S., 2007. Reduced TOR signaling extends chronological life span via increased respiration and upregulation of mitochondrial gene expression. Cell Metab. 5, 265–277.
- Bonkowski, M.S., Rocha, J.S., Masternak, M.M., Al Regaiey, K.A., Bartke, A., 2006. Targeted disruption of growth hormone receptor interferes with the beneficial actions of calorie restriction. Proc. Natl. Acad. Sci. USA 103, 7901–7905.
- Bonkowski, M.S., Dominici, F.P., Arum, O., Rocha, J.S., Al Regaiey, K.A., Westbrook, R., Spong, A., Panici, J., Masternak, M.M., Kopchick, J.J., Bartke, A., 2009. Disruption of growth hormone receptor prevents calorie restriction from improving insulin action and longevity. PLoS One 4, e4567.
- Bratic, I., Trifunovic, A., 2010. Mitochondrial energy metabolism and ageing. Biochim. Biophys. Acta.
- Brooks, N.L., Trent, C.M., Raetzsch, C.F., Flurkey, K., Boysen, G., Perfetti, M.T., Jeong, Y.C., Klebanov, S., Patel, K.B., Khodush, V.R., Kupper, L.L., Carling, D., Swenberg, J.A., Harrison, D.E., Combs, T.P., 2007. Low utilization of circulating glucose after food withdrawal in Snell dwarf mice. J. Biol. Chem. 282, 35069–35077.
- Brown-Borg, H.M., Borg, K.E., Meliska, C.J., Bartke, A., 1996. Dwarf mice and the ageing process. Nature 384, 33.
- Brown-Borg, H.M., Rakoczy, S.G., Romanick, M.A., Kennedy, M.A., 2002. Effects of growth hormone and insulin-like growth factor-1 on hepatocyte antioxidative enzymes. Exp. Biol. Med. (Maywood) 227, 94–104.
- Brüning, J.C., Michael, M.D., Winnay, J.N., Hayashi, T., Hörsch, D., Accili, D., Goodyear, L.J., Kahn, C.R., 1998. A muscle-specific insulin receptor knockout exhibits features of the metabolic syndrome of NIDDM without altering glucose tolerance. Mol. Cell. 2, 559–569.
- Calabrese, E.J., Bachmann, K.A., Bailer, A.J., Bolger, P.M., Borak, J., Cai, L., Cedergreen, N., Cherian, M.G., Chiueh, C.C., Clarkson, T.W., Cook, R.R., Diamond, D.M., Doolittle, D.J., Dorato, M.A., Duke, S.O., Feinendegen, L., Gardner, D.E., Hart, R.W., Hastings, K.L., Hayes, A.W., Hoffmann, G.R., Ives, J.A., Jaworowski, Z., Johnson, T.E., Jonas, W.B., Kaminski, N.E., Keller, J.G., Klaunig, J.E., Knudsen, T.B., Kozumbo, W.J., Lettieri, T., Liu, S.Z., Maisseu, A., Maynard, K.I., Masoro, E.J., McClellan, R.O., Mehendale, H.M., Mothersill, C., Newlin, D.B., Nigg, H.N., Oehme, F.W., Phalen, R.F., Philbert, M.A., Rattan, S.I., Riviere, J.E., Rodricks, J., Sapolsky, R.M., Scott, B.R., Seymour, C., Sinclair, D.A., Smith-Sonneborn, J., Snow, E.T., Spear, L., Stevenson, D.E., Thomas, Y., Tubiana, M., Williams, G.M., Mattson, M.P., 2007. Biological stress response terminology: integrating the concepts of adaptive response and preconditioning stress within a hormetic dose-response framework. Toxicol. Appl. Pharmacol. 222, 122–128.
- Chandel, N.S., Budinger, G.R., 2007. The cellular basis for diverse responses to oxygen. Free Radic. Biol. Med. 42, 165–174.
- Chevion, S., Moran, D.S., Heled, Y., Shani, Y., Regev, G., Abbou, B., Berenshtein, E., Stadtman, E.R., Epstein, Y., 2003. Plasma antioxidant status and cell injury after severe physical exercise. Proc. Natl. Acad. Sci. USA 100, 5119–5123.
- Clancy, D.J., Gems, D., Hafen, E., Leevers, S.J., Partridge, L., 2002. Dietary restriction in long-lived dwarf flies. Science 296, 319.
- Cohen, E., Paulsson, J.F., Blinder, P., Burstyn-Cohen, T., Du, D., Estepa, G., Adame, A., Pham, H.M., Holzenberger, M., Kelly, J.W., Masliah, E., Dillin, A., 2009. Reduced IGF-1 signaling delays age-associated proteotoxicity in mice. Cell 139, 1157-1169
- Colman, R.J., Anderson, R.M., Johnson, S.C., Kastman, E.K., Kosmatka, K.J., Beasley, T.M., Allison, D.B., Cruzen, C., Simmons, H.A., Kemnitz, J.W., Weindruch, R., 2009. Caloric restriction delays disease onset and mortality in rhesus monkeys. Science 325, 201–204.
- Connor, K.M., Subbaram, S., Regan, K.J., Nelson, K.K., Mazurkiewicz, J.E., Bartholomew, P.J., Aplin, A.E., Tai, Y.T., Aguirre-Ghiso, J., Flores, S.C., Melendez, J.A., 2005. Mitochondrial H₂O₂ regulates the angiogenic phenotype via PTEN oxidation. J. Biol. Chem. 280, 16916–16924.
- Cook, N.R., Albert, C.M., Gaziano, J.M., Zaharris, E., MacFadyen, J., Danielson, E., Buring, J.E., Manson, J.E., 2007. A randomized factorial trial of vitamins C and E

- and beta carotene in the secondary prevention of cardiovascular events in women: results from the women's antioxidant cardiovascular study. Arch. Intern. Med. 167, 1610–1618.
- Czernichow, S., Bertrais, S., Blacher, J., Galan, P., Briancon, S., Favier, A., Safar, M., Hercberg, S., 2005. Effect of supplementation with antioxidants upon long-term risk of hypertension in the SU.VI.MAX study: association with plasma antioxidant levels. J. Hypertens. 23, 2013–2018.
- Czernichow, S., Couthouis, A., Bertrais, S., Vergnaud, A.C., Dauchet, L., Galan, P., Hercberg, S., 2006. Antioxidant supplementation does not affect fasting plasma glucose in the Supplementation with Antioxidant Vitamins and Minerals (SU.VI.MAX) study in France association with dietary intake and plasma concentrations. Am. J. Clin. Nutr. 84, 395–399.
- Dai, D.F., Santana, L.F., Vermulst, M., Tomazela, D.M., Emond, M.J., MacCoss, M.J., Gollahon, K., Martin, G.M., Loeb, L.A., Ladiges, W.C., Rabinovitch, P.S., 2009. Overexpression of catalase targeted to mitochondria attenuates murine cardiac aging. Circulation 119, 2789–2797.
- Davies, K.J., Quintanilha, A.T., Brooks, G.A., Packer, L., 1982. Free radicals and tissue damage produced by exercise. Biochem. Biophys. Res. Commun. 107, 1198–1205.
- De, A.K., Chipalkatti, S., Aiyar, A.S., 1983. Some biochemical parameters of ageing in relation to dietary protein. Mech. Ageing Dev. 21, 37–48.
- DeFronzo, R.A., 1981. Glucose intolerance and aging. Diabetes Care 4, 493-501.
- Dong, M.Q., Venable, J.D., Au, N., Xu, T., Park, S.K., Cociorva, D., Johnson, J.R., Dillin, A., Yates 3rd, J.R., 2007. Quantitative mass spectrometry identifies insulin signaling targets in *C. elegans*. Science 317, 660–663.
- Doonan, R., McElwee, J.J., Matthijssens, F., Walker, G.A., Houthoofd, K., Back, P., Matscheski, A., Vanfleteren, J.R., Gems, D., 2008. Against the oxidative damage theory of aging: superoxide dismutases protect against oxidative stress but have little or no effect on life span in *Caenorhabditis elegans*. Genes Dev. 22, 3236–3241.
- Duan, W., Mattson, M.P., 1999. Dietary restriction and 2-deoxyglucose administration improve behavioral outcome and reduce degeneration of dopaminergic neurons in models of Parkinson's disease. J. Neurosci. Res. 57, 195–206.
- Finley, L.W., Haigis, M.C., 2009. The coordination of nuclear and mitochondrial communication during aging and calorie restriction. Ageing Res. Rev. 8, 173–188.
- Fontana, L., Klein, S., 2007. Aging, adiposity, and calorie restriction. JAMA 297, 986–994.
- Fontana, L., Meyer, T.E., Klein, S., Holloszy, J.O., 2004. Long-term calorie restriction is highly effective in reducing the risk for atherosclerosis in humans. Proc. Natl. Acad. Sci. USA 101, 6659–6663.
- Friedman, D.B., Johnson, T.E., 1988. A mutation in the age-1 gene in *Caenorhabditis elegans* lengthens life and reduces hermaphrodite fertility. Genetics 118, 75–86.
- Fukui, H., Moraes, C.T., 2008. The mitochondrial impairment, oxidative stress and neurodegeneration connection: reality or just an attractive hypothesis? Trends Neurosci. 31, 251–256.
- Garriga-Canut, M., Schoenike, B., Qazi, R., Bergendahl, K., Daley, T.J., Pfender, R.M., Morrison, J.F., Ockuly, J., Stafstrom, C., Sutula, T., Roopra, A., 2006. 2-Deoxy-pglucose reduces epilepsy progression by NRSF-CtBP-dependent metabolic regulation of chromatin structure. Nat. Neurosci. 9, 1382–1387.
- Gomez-Cabrera, M.C., Domenech, E., Romagnoli, M., Arduini, A., Borras, C., Pallardo, F.V., Sastre, J., Vina, J., 2008. Oral administration of vitamin C decreases muscle mitochondrial biogenesis and hampers training-induced adaptations in endurance performance. Am. J. Clin. Nutr. 87, 142–149.
- Grandison, R.C., Piper, M.D., Partridge, L., 2009. Amino-acid imbalance explains extension of lifespan by dietary restriction in Drosophila. Nature 462, 1061–1064
- Gredilla, R., Barja, G., 2005. Minireview: the role of oxidative stress in relation to caloric restriction and longevity. Endocrinology 146, 3713–3717.
- Greenberg, E.R., Baron, J.A., Tosteson, T.D., Freeman Jr., D.H., Beck, G.J., Bond, J.H., Colacchio, T.A., Coller, J.A., Frankl, H.D., Haile, R.W., et al., 1994. A clinical trial of antioxidant vitamins to prevent colorectal adenoma. Polyp prevention study group. N. Engl. J. Med. 331, 141–147.
- Greer, E.L., Dowlatshahi, D., Banko, M.R., Villen, J., Hoang, K., Blanchard, D., Gygi, S.P.,
 Brunet, A., 2007. An AMPK-FOXO pathway mediates longevity induced by a
 novel method of dietary restriction in *C. elegans*. Curr. Biol. 17. 1646-1656.
- Guarente, L., 2006. Sirtuins as potential targets for metabolic syndrome. Nature 444, 868–874.
- Guzy, R.D., Schumacker, P.T., 2006. Oxygen sensing by mitochondria at complex III: the paradox of increased reactive oxygen species during hypoxia. Exp. Physiol. 91, 807–819.
- Guzy, R.D., Hoyos, B., Robin, E., Chen, H., Liu, L., Mansfield, K.D., Simon, M.C., Hammerling, U., Schumacker, P.T., 2005. Mitochondrial complex III is required for hypoxia-induced ROS production and cellular oxygen sensing. Cell Metab. 1, 401–408
- Gwinn, D.M., Shackelford, D.B., Egan, D.F., Mihaylova, M.M., Mery, A., Vasquez, D.S., Turk, B.E., Shaw, R.J., 2008. AMPK phosphorylation of raptor mediates a metabolic checkpoint. Mol. Cell. 30, 214–226.
- Hardie, D.G., Hawley, S.A., Scott, J.W., 2006. AMP-activated protein kinase: development of the energy sensor concept. J. Physiol. 574, 7–15.
- Harman, D., 1956. Aging: a theory based on free radical and radiation chemistry. J. Gerontol. 11, 298–300.
- Harrington, L.A., Harley, C.B., 1988. Effect of vitamin E on lifespan and reproduction in *Caenorhabditis elegans*. Mech. Ageing Dev. 43, 71–78.

- Heart Protection Study Collaborative Group, 2002. MRC/BHF Heart Protection Study of antioxidant vitamin supplementation in 20,536 high-risk individuals: a randomised placebo-controlled trial. Lancet 360, 23-33.
- Heidler, T., Hartwig, K., Daniel, H., Wenzel, U., 2009. *Caenorhabditis elegans* lifespan extension caused by treatment with an orally active ROS-generator is dependent on DAF-16 and SIR-2.1. Biogerontology.
- Heilbronn, L.K., Ravussin, E., 2003. Calorie restriction and aging: review of the literature and implications for studies in humans. Am. J. Clin. Nutr. 78, 361–369.
- Heilbronn, L.K., de Jonge, L., Frisard, M.I., DeLany, J.P., Larson-Meyer, D.E., Rood, J., Nguyen, T., Martin, C.K., Volaufova, J., Most, M.M., Greenway, F.L., Smith, S.R., Deutsch, W.A., Williamson, D.A., Ravussin, E., 2006. Effect of 6-month calorie restriction on biomarkers of longevity, metabolic adaptation, and oxidative stress in overweight individuals: a randomized controlled trial. JAMA 295, 1539–1548.
- Hercberg, S., Ezzedine, K., Guinot, C., Preziosi, P., Galan, P., Bertrais, S., Estaquio, C., Briancon, S., Favier, A., Latreille, J., Malvy, D., 2007. Antioxidant supplementation increases the risk of skin cancers in women but not in men. J. Nutr. 137, 2098–2105.
- Hession, M., Rolland, C., Kulkarni, U., Wise, A., Broom, J., 2009. Systematic review of randomized controlled trials of low-carbohydrate vs. low-fat/low-calorie diets in the management of obesity and its comorbidities. Obes. Rev. 10, 36–50.
- Higuchi, M., Cartier, L.J., Chen, M., Holloszy, J.O., 1985. Superoxide dismutase and catalase in skeletal muscle: adaptive response to exercise. J. Gerontol. 40, 281–286.
- Holzenberger, M., Dupont, J., Ducos, B., Leneuve, P., Geloen, A., Even, P.C., Cervera, P., Le Bouc, Y., 2003. IGF-1 receptor regulates lifespan and resistance to oxidative stress in mice. Nature 421, 182–187.
- Honda, Y., Honda, S., 1999. The daf-2 gene network for longevity regulates oxidative stress resistance and Mn-superoxide dismutase gene expression in *Caenorhabditis elegans*. FASEB J. 13, 1385–1393.
- Houthoofd, K., Braeckman, B.P., Lenaerts, I., Brys, K., De Vreese, A., Van Eygen, S., Vanfleteren, J.R., 2002. No reduction of metabolic rate in food restricted Caenorhabditis elegans. Exp. Gerontol. 37, 1359–1369.
- Houthoofd, K., Braeckman, B.P., Johnson, T.E., Vanfleteren, J.R., 2003. Life extension via dietary restriction is independent of the Ins/IGF-1 signalling pathway in *Caenorhabditis elegans*. Exp. Gerontol. 38, 947–954.
- Houthoofd, K., Fidalgo, M.A., Hoogewijs, D., Braeckman, B.P., Lenaerts, I., Brys, K., Matthijssens, F., De Vreese, A., Van Eygen, S., Munoz, M.J., Vanfleteren, J.R., 2005. Metabolism, physiology and stress defense in three aging Ins/IGF-1 mutants of the nematode *Caenorhabditis elegans*. Aging Cell 4, 87–95.
- Huang, T.T., Carlson, E.J., Gillespie, A.M., Shi, Y., Epstein, C.J., 2000. Ubiquitous overexpression of CuZn superoxide dismutase does not extend life span in mice. J. Gerontol. A Biol. Sci. Med. Sci. 55, B5–B9.
- Huang, T.T., Naeemuddin, M., Elchuri, S., Yamaguchi, M., Kozy, H.M., Carlson, E.J., Epstein, C.J., 2006. Genetic modifiers of the phenotype of mice deficient in mitochondrial superoxide dismutase. Hum. Mol. Genet. 15, 1187–1194.
- Ingram, D.K., Anson, R.M., de Cabo, R., Mamczarz, J., Zhu, M., Mattison, J., Lane, M.A., Roth, G.S., 2004. Development of calorie restriction mimetics as a prolongevity strategy. Ann. NY. Acad. Sci. 1019, 412–423.
- Ingram, D.K., Roth, G.S., Lane, M.A., Ottinger, M.A., Zou, S., de Cabo, R., Mattison, J.A., 2006a. The potential for dietary restriction to increase longevity in humans: extrapolation from monkey studies. Biogerontology 7, 143–148.
- Ingram, D.K., Zhu, M., Mamczarz, J., Zou, S., Lane, M.A., Roth, G.S., deCabo, R., 2006b. Calorie restriction mimetics: an emerging research field. Aging Cell 5, 97–108.
- Ishii, N., Senoo-Matsuda, N., Miyake, K., Yasuda, K., Ishii, T., Hartman, P.S., Furukawa, S., 2004. Coenzyme Q10 can prolong *C. elegans* lifespan by lowering oxidative stress. Mech. Ageing Dev. 125, 41–46.
- Jang, Y.C., van Remmen, H., 2009. The mitochondrial theory of aging: insight from transgenic and knockout mouse models. Exp. Gerontol. 44, 256–260.
- Jang, Y.C., Perez, V.I., Song, W., Lustgarten, M.S., Salmon, A.B., Mele, J., Qi, W., Liu, Y., Liang, H., Chaudhuri, A., Ikeno, Y., Epstein, C.J., Van Remmen, H., Richardson, A., 2009. Overexpression of Mn superoxide dismutase does not increase life span in mice. J. Gerontol. A Biol. Sci. Med. Sci. 64, 1114–1125.
- Kaeberlein, M., Kirkland, K.T., Fields, S., Kennedy, B.K., 2004. Sir2-independent life span extension by calorie restriction in yeast. PLoS Biol. 2, E296.
- Kaeberlein, M., Hu, D., Kerr, E.O., Tsuchiya, M., Westman, E.A., Dang, N., Fields, S., Kennedy, B.K., 2005. Increased life span due to calorie restriction in respiratorydeficient yeast. PLoS Genet. 1, e69.
- Kaelin Jr., W.G., 2005. ROS: really involved in oxygen sensing. Cell Metab. 1, 357–358.
- Kahn, C.R., 1994. Banting lecture: insulin action, diabetogenes, and the cause of type II diabetes. Diabetes 43, 1066–1084.
- Kappeler, L., De Magalhaes Filho, C.M., Dupont, J., Leneuve, P., Cervera, P., Perin, L., Loudes, C., Blaise, A., Klein, R., Epelbaum, J., Le Bouc, Y., Holzenberger, M., 2008. Brain IGF-1 receptors control mammalian growth and lifespan through a neuroendocrine mechanism. PLoS Biol. 6, e254.
- Kataja-Tuomola, M., Sundell, J.R., Mannisto, S., Virtanen, M.J., Kontto, J., Albanes, D., Virtamo, J., 2008. Effect of alpha-tocopherol and beta-carotene supplementation on the incidence of type 2 diabetes. Diabetologia 51, 47–53.
- Katic, M., Kennedy, A.R., Leykin, I., Norris, A., McGettrick, A., Gesta, S., Russell, S.J., Bluher, M., Maratos-Flier, E., Kahn, C.R., 2007. Mitochondrial gene expression and increased oxidative metabolism: role in increased lifespan of fat-specific insulin receptor knock-out mice. Aging Cell 6, 827–839.
- Katsiki, N., Manes, C., 2009. Is there a role for supplemented antioxidants in the prevention of atherosclerosis? Clin. Nutr. 28, 3–9.

- Keaney, M., Gems, D., 2003. No increase in lifespan in *Caenorhabditis elegans* upon treatment with the superoxide dismutase mimetic EUK-8. Free Radic. Biol. Med. 34, 277–282.
- Kenyon, C., Chang, J., Gensch, E., Rudner, A., Tabtiang, R., 1993. A *C. elegans* mutant that lives twice as long as wild type. Nature 366, 461–464.
- Kharade, S.V., Mittal, N., Das, S.P., Sinha, P., Roy, N., 2005. Mrg19 depletion increases S. cerevisiae lifespan by augmenting ROS defence. FEBS Lett. 579, 6809–6813.
- Kim, J., Takahashi, M., Shimizu, T., Shirasawa, T., Kajita, M., Kanayama, A., Miyamoto, Y., 2008. Effects of a potent antioxidant, platinum nanoparticle, on the lifespan of *Caenorhabditis elegans*. Mech. Ageing Dev. 129, 322–331.
- Kimura, K.D., Tissenbaum, H.A., Liu, Y., Ruvkun, G., 1997. daf-2, an insulin receptorlike gene that regulates longevity and diapause in Caenorhabditis elegans. Science 277, 942–946.
- Koizumi, A., Weindruch, R., Walford, R.L., 1987. Influences of dietary restriction and age on liver enzyme activities and lipid peroxidation in mice. J. Nutr. 117, 361–367
- Kotani, K., Peroni, O.D., Minokoshi, Y., Boss, O., Kahn, B.B., 2004. GLUT4 glucose transporter deficiency increases hepatic lipid production and peripheral lipid utilization. J. Clin. Invest. 114, 1666–1675.
- Lakowski, B., Hekimi, S., 1998. The genetics of caloric restriction in *Caenorhabditis elegans*. Proc. Natl. Acad. Sci. USA 95, 13091–13096.
- Lane, M.A., 1998. 2-Deoxy-D-glucose feeding in rats mimics physiologic effects of calorie restriction. J. Anti-Aging Med. 1, 327–336.
- Lanza, I.R., Short, D.K., Short, K.R., Raghavakaimal, S., Basu, R., Joyner, M.J., McConnell, J.P., Nair, K.S., 2008. Endurance exercise as a countermeasure for aging. Diabetes 57, 2933–2942.
- Lapointe, J., Stepanyan, Z., Bigras, E., Hekimi, S., 2009. Reversal of the mitochondrial phenotype and slow development of oxidative biomarkers of aging in long-lived Mclk1+/- mice. J. Biol. Chem. 284, 20364–20374.
- Lawenda, B.D., Kelly, K.M., Ladas, E.J., Sagar, S.M., Vickers, A., Blumberg, J.B., 2008. Should supplemental antioxidant administration be avoided during chemotherapy and radiation therapy? J. Natl. Cancer Inst. 100, 773–783.
- Lee, S.J., Murphy, C.T., Kenyon, C., 2009. Glucose shortens the life span of *C. elegans* by downregulating DAF-16/FOXO activity and aquaporin gene expression. Cell Metab. 10, 379–391.
- Lin, S.J., Defossez, P.A., Guarente, L., 2000. Requirement of NAD and SIR2 for lifespan extension by calorie restriction in Saccharomyces cerevisiae. Science 289, 2126–2128.
- Lin, S.J., Kaeberlein, M., Andalis, A.A., Sturtz, L.A., Defossez, P.A., Culotta, V.C., Fink, G.R., Guarente, L., 2002. Calorie restriction extends Saccharomyces cerevisiae lifespan by increasing respiration. Nature 418, 344–348.
- Lin, J., Cook, N.R., Albert, C., Zaharris, E., Gaziano, J.M., Van Denburgh, M., Buring, J.E., Manson, J.E., 2009. Vitamins C and E and beta-carotene supplementation and cancer risk: a randomized controlled trial. J. Natl. Cancer Inst. 101, 14–23.
- Lindsted, K.D., Tonstad, S., Kuzma, J.W., 1991. Self-report of physical activity and patterns of mortality in seventh-day adventist men. J. Clin. Epidemiol. 44, 355–364.
- Lippman, S.M., Klein, E.A., Goodman, P.J., Lucia, M.S., Thompson, I.M., Ford, L.G., Parnes, H.L., Minasian, L.M., Gaziano, J.M., Hartline, J.A., Parsons, J.K., Bearden 3rd, J.D., Crawford, E.D., Goodman, G.E., Claudio, J., Winquist, E., Cook, E.D., Karp, D.D., Walther, P., Lieber, M.M., Kristal, A.R., Darke, A.K., Arnold, K.B., Ganz, P.A., Santella, R.M., Albanes, D., Taylor, P.R., Probstfield, J.L., Jagpal, T.J., Crowley, J.J., Meyskens Jr., F.L., Baker, L.H., Coltman Jr., C.A., 2009. Effect of selenium and vitamin E on risk of prostate cancer and other cancers: the selenium and vitamin e cancer prevention trial (SELECT). JAMA 301, 39–51.
- Lithgow, G.J., White, T.M., Melov, S., Johnson, T.E., 1995. Thermotolerance and extended life-span conferred by single-gene mutations and induced by thermal stress. Proc. Natl. Acad. Sci. USA 92, 7540–7544.

 Liu, S., Ajani, U., Chae, C., Hennekens, C., Buring, J.E., Manson, J.E., 1999. Long-term
- Liu, S., Ajani, U., Chae, C., Hennekens, C., Buring, J.E., Manson, J.E., 1999. Long-term beta-carotene supplementation and risk of type 2 diabetes mellitus: a randomized controlled trial. JAMA 282, 1073–1075.
- Loh, K., Deng, H., Fukushima, A., Cai, X., Boivin, B., Galic, S., Bruce, C., Shields, B.J., Skiba, B., Ooms, L.M., Stepto, N., Wu, B., Mitchell, C.A., Tonks, N.K., Watt, M.J., Febbraio, M.A., Crack, P.J., Andrikopoulos, S., Tiganis, T., 2009. Reactive oxygen species enhance insulin sensitivity. Cell Metab. 10, 260–272.
- Lonn, E., Bosch, J., Yusuf, S., Sheridan, P., Pogue, J., Arnold, J.M., Ross, C., Arnold, A., Sleight, P., Probstfield, J., Dagenais, G.R., 2005. Effects of long-term vitamin E supplementation on cardiovascular events and cancer: a randomized controlled trial. JAMA 293, 1338–1347.
- Lopez-Lluch, G., Hunt, N., Jones, B., Zhu, M., Jamieson, H., Hilmer, S., Cascajo, M.V., Allard, J., Ingram, D.K., Navas, P., de Cabo, R., 2006. Calorie restriction induces mitochondrial biogenesis and bioenergetic efficiency. Proc. Natl. Acad. Sci. USA 103, 1768–1773.
- Mair, W., Piper, M.D., Partridge, L., 2005. Calories do not explain extension of life span by dietary restriction in Drosophila. PLoS Biol. 3, e223.
- Manini, T.M., Everhart, J.E., Patel, K.V., Schoeller, D.A., Colbert, L.H., Visser, M., Tylavsky, F., Bauer, D.C., Goodpaster, B.H., Harris, T.B., 2006. Daily activity energy expenditure and mortality among older adults. JAMA 296, 171–179.
- Masoro, E.J., 1998b. Influence of caloric intake on aging and on the response to stressors. J. Toxicol. Environ. Health B Crit. Rev. 1, 243–257.
- Masoro, E.J., 1998a. Hormesis and the antiaging action of dietary restriction. Exp. Gerontol. 33, 61-66.
- Masoro, E.J., 2000. Caloric restriction and aging: an update. Exp. Gerontol. 35, 299–305.
- Masoro, E.J., Yu, B.P., Bertrand, H.A., 1982. Action of food restriction in delaying the aging process. Proc. Natl. Acad. Sci. USA 79, 4239–4241.

- McCarter, R., Mejia, W., Ikeno, Y., Monnier, V., Kewitt, K., Gibbs, M., McMahan, A., Strong, R., 2007. Plasma glucose and the action of calorie restriction on aging. J. Gerontol. A Biol. Sci. Med. Sci. 62, 1059–1070.
- McCay, C.M., Crowel, M.F., Maynard, L.A., 1935. The effect of retarded growth upon the length of the life span and upon ultimate body size. J. Nutr. 10, 63–79.
- Meissner, B., Boll, M., Daniel, H., Baumeister, R., 2004. Deletion of the intestinal peptide transporter affects insulin and TOR signaling in Caenorhabditis elegans. J. Biol. Chem. 279, 36739–36745.
- Melov, S., Coskun, P., Patel, M., Tuinstra, R., Cottrell, B., Jun, A.S., Zastawny, T.H., Dizdaroglu, M., Goodman, S.I., Huang, T.T., Miziorko, H., Epstein, C.J., Wallace, D.C., 1999. Mitochondrial disease in superoxide dismutase 2 mutant mice. Proc. Natl. Acad. Sci. USA 96, 846–851.
- Melov, S., Ravenscroft, J., Malik, S., Gill, M.S., Walker, D.W., Clayton, P.E., Wallace, D.C., Malfroy, B., Doctrow, S.R., Lithgow, G.J., 2000. Extension of life-span with superoxide dismutase/catalase mimetics. Science 289, 1567–1569.
- Miller, R.A., Buehner, G., Chang, Y., Harper, J.M., Sigler, R., Smith-Wheelock, M., 2005. Methionine-deficient diet extends mouse lifespan, slows immune and lens aging, alters glucose, T4, IGF-I and insulin levels, and increases hepatocyte MIF levels and stress resistance. Aging Cell 4, 119–125.
- Min, K.J., Tatar, M., 2006. Restriction of amino acids extends lifespan in *Drosophila melanogaster*. Mech. Ageing Dev. 127, 643-646.
- Min, K.J., Yamamoto, R., Buch, S., Pankratz, M., Tatar, M., 2008. Drosophila lifespan control by dietary restriction independent of insulin-like signaling. Aging Cell 7, 199–206.
- Miwa, S., Riyahi, K., Partridge, L., Brand, M.D., 2004. Lack of correlation between mitochondrial reactive oxygen species production and life span in Drosophila. Ann. NY. Acad. Sci. 1019, 388–391.
- Moskovitz, J., Bar-Noy, S., Williams, W.M., Requena, J., Berlett, B.S., Stadtman, E.R., 2001. Methionine sulfoxide reductase (MsrA) is a regulator of antioxidant defense and lifespan in mammals. Proc. Natl. Acad. Sci. USA 98, 12920–12925.
- Murphy, C.T., McCarroll, S.A., Bargmann, C.I., Fraser, A., Kamath, R.S., Ahringer, J., Li, H., Kenyon, C., 2003. Genes that act downstream of DAF-16 to influence the lifespan of *Caenorhabditis elegans*. Nature 424, 277–283.
- Myung, S.K., Kim, Y., Ju, W., Choi, H.J., Bae, W.K., 2010. Effects of antioxidant supplements on cancer prevention: meta-analysis of randomized controlled trials. Ann. Oncol. 21, 166–179.
- Narasimhan, S.D., Yen, K., Tissenbaum, H.A., 2009. Converging pathways in lifespan regulation. Curr. Biol. 19, R657–R666.
- Nazarewicz, R.R., Ziolkowski, W., Vaccaro, P.S., Ghafourifar, P., 2007. Effect of shortterm ketogenic diet on redox status of human blood. Rejuvenation Res. 10, 435– 440
- Nisoli, E., Tonello, C., Cardile, A., Cozzi, V., Bracale, R., Tedesco, L., Falcone, S., Valerio, A., Cantoni, O., Clementi, E., Moncada, S., Carruba, M.O., 2005. Calorie restriction promotes mitochondrial biogenesis by inducing the expression of eNOS. Science 310, 314–317.
- Nordmann, A.J., Nordmann, A., Briel, M., Keller, U., Yancy Jr., W.S., Brehm, B.J., Bucher, H.C., 2006. Effects of low-carbohydrate vs low-fat diets on weight loss and cardiovascular risk factors: a meta-analysis of randomized controlled trials. Arch. Intern. Med. 166, 285–293.
- Orr, W.C., Sohal, R.S., 1994. Extension of life-span by overexpression of superoxide dismutase and catalase in *Drosophila melanogaster*. Science 263, 1128–1130.
- Owusu-Ansah, E., Yavari, A., Mandal, S., Banerjee, U., 2008. Distinct mitochondrial retrograde signals control the G1-S cell cycle checkpoint. Nat. Genet. 40, 356–361.
- Parkes, T.L., Elia, A.J., Dickinson, D., Hilliker, A.J., Phillips, J.P., Boulianne, G.L., 1998. Extension of Drosophila lifespan by overexpression of human SOD1 in motorneurons. Nat. Genet. 19, 171–174.
- Pawlikowska, L., Hu, D., Huntsman, S., Sung, A., Chu, C., Chen, J., Joyner, A., Schork, N.J., Hsueh, W.C., Reiner, A.P., Psaty, B.M., Atzmon, G., Barzilai, N., Cummings, S.R., Browner, W.S., Kwok, P.Y., Ziv, E., 2009. Association of common genetic variation in the insulin/IGF1 signaling pathway with human longevity. Aging Cell 8, 460–472.
- Pearl, R., 1928. The Rate of Living. Being an Account of Some Experimental Studies on the Biology of Life Duration. Alfred Knopf, New York.
- Pendergrass, W.R., Li, Y., Jiang, D., Wolf, N.S., 1993. Decrease in cellular replicative potential in "giant" mice transfected with the bovine growth hormone gene correlates to shortened life span. J. Cell. Physiol. 156, 96–103.
- Phillips, J.P., Campbell, S.D., Michaud, D., Charbonneau, M., Hilliker, A.J., 1989. Null mutation of copper/zinc superoxide dismutase in Drosophila confers hypersensitivity to paraquat and reduced longevity. Proc. Natl. Acad. Sci. USA 86, 2761–2765.
- Pieri, C., Falasca, M., Marcheselli, F., Moroni, F., Recchioni, R., Marmocchi, F., Lupidi, G., 1992. Food restriction in female Wistar rats: V. Lipid peroxidation and antioxidant enzymes in the liver. Arch. Gerontol. Geriatr. 14, 93–99.
- Piper, M.D., Bartke, A., 2008. Diet and aging. Cell Metab. 8, 99-104.
- Piper, P.W., Harris, N.L., MacLean, M., 2006. Preadaptation to efficient respiratory maintenance is essential both for maximal longevity and the retention of replicative potential in chronologically ageing yeast. Mech. Ageing Dev. 127, 733– 740.
- Powers, S.K., Jackson, M.J., 2008. Exercise-induced oxidative stress: cellular mechanisms and impact on muscle force production. Physiol. Rev. 88, 1243– 1276.
- Powers 3rd, R.W., Kaeberlein, M., Caldwell, S.D., Kennedy, B.K., Fields, S., 2006. Extension of chronological life span in yeast by decreased TOR pathway signaling. Genes Dev. 20, 174–184.

- Pun, P.B., Gruber, J., Tang, S.Y., Schaffer, S., Ong, R.L., Fong, S., Ng, L.F., Cheah, I., Halliwell, B., 2010. Ageing in nematodes: do antioxidants extend lifespan in Caenorhabditis elegans? Biogerontology 11, 17–30.
- Quarrie, J.K., Riabowol, K.T., 2004. Murine models of life span extension. Sci. Aging Knowledge Environ. re5.
- Quick, K.L., Ali, S.S., Arch, R., Xiong, C., Wozniak, D., Dugan, L.L., 2008. A carboxyfullerene SOD mimetic improves cognition and extends the lifespan of mice. Neurobiol. Aging 29, 117–128.
- Ran, Q., Liang, H., Ikeno, Y., Qi, W., Prolla, T.A., Roberts 2nd, L.J., Wolf, N., Van Remmen, H., Richardson, A., 2007. Reduction in glutathione peroxidase 4 increases life span through increased sensitivity to apoptosis. J. Gerontol. A Biol. Sci. Med. Sci. 62, 932–942.
- Rao, G., Xia, E., Nadakavukaren, M.J., Richardson, A., 1990. Effect of dietary restriction on the age-dependent changes in the expression of antioxidant enzymes in rat liver. J. Nutr. 120, 602–609.
- Rautalahti, M.T., Virtamo, J.R., Taylor, P.R., Heinonen, O.P., Albanes, D., Haukka, J.K., Edwards, B.K., Karkkainen, P.A., Stolzenberg-Solomon, R.Z., Huttunen, J., 1999. The effects of supplementation with alpha-tocopherol and beta-carotene on the incidence and mortality of carcinoma of the pancreas in a randomized, controlled trial. Cancer 86, 37–42.
- Rhee, S.G., Chang, T.S., Bae, Y.S., Lee, S.R., Kang, S.W., 2003. Cellular regulation by hydrogen peroxide. J. Am. Soc. Nephrol. 14, S211–S215.
- Ristow, M., 2006. Oxidative metabolism in cancer growth. Curr. Opin. Clin. Nutr. Metabol. 9, 339–345.
- Ristow, M., Zarse, K., Oberbach, A., Klöting, N., Birringer, M., Kiehntopf, M., Stumvoll, M., Kahn, C.R., Blüher, M., 2009. Antioxidants prevent health-promoting effects of physical exercise in humans. Proc. Nat. Acad. Sci. USA 106, 8665–8670.
- Roux, A.E., Leroux, A., Alaamery, M.A., Hoffman, C.S., Chartrand, P., Ferbeyre, G., Rokeach, L.A., 2009. Pro-aging effects of glucose signaling through a G proteincoupled glucose receptor in fission yeast. PLoS Genet. 5, e1000408.
- Ruan, H., Tang, X.D., Chen, M.L., Joiner, M.L., Sun, G., Brot, N., Weissbach, H., Heinemann, S.H., Iverson, L., Wu, C.F., Hoshi, T., 2002. High-quality life extension by the enzyme peptide methionine sulfoxide reductase. Proc. Natl. Acad. Sci. USA 99, 2748–2753.
- Rubner, M., 1908. III. Das Wachstumsproblem und die Lebensdauer des Menschen und einiger Säugetiere vom energetischen Standpunkt aus betrachtet. In: Rubner, M. (Ed.), Das Problem der Lebensdauer und seine Beziehungen zum Wachstum und der Ernährung. R. Oldenbourg, Munich and Berlin, pp. 127–208.
- Russell, S.J., Kahn, C.R., 2007. Endocrine regulation of ageing. Nat. Rev. Mol. Cell. Biol. 8, 681–691.
- Sacco, M., Pellegrini, F., Roncaglioni, M.C., Avanzini, F., Tognoni, G., Nicolucci, A., 2003. Primary prevention of cardiovascular events with low-dose aspirin and vitamin E in type 2 diabetic patients: results of the Primary Prevention Project (PPP) trial. Diabetes Care 26, 3264–3272.
- Schlotterer, A., Kukudov, G., Bozorgmehr, F., Hutter, H., Du, X., Oikonomou, D., Ibrahim, Y., Pfisterer, F., Rabbani, N., Thornalley, P., Sayed, A., Fleming, T., Humpert, P., Schwenger, V., Zeier, M., Hamann, A., Stern, D., Brownlee, M., Bierhaus, A., Nawroth, P., Morcos, M., 2009. C. elegans as model for the study of high glucose mediated lifespan reduction. Diabetes 58, 2450–2456.
- Sesso, H.D., Buring, J.E., Christen, W.G., Kurth, T., Belanger, C., MacFadyen, J., Bubes, V., Manson, J.E., Glynn, R.J., Gaziano, J.M., 2008. Vitamins E and C in the prevention of cardiovascular disease in men: the Physicians' Health Study II randomized controlled trial. JAMA 300, 2123–2133.
- Schulz, T.J., Zarse, K., Voigt, A., Urban, N., Birringer, M., Ristow, M., 2007. Glucose restriction extends *Caenorhabditis elegans* life span by inducing mitochondrial respiration and increasing oxidative stress. Cell Metab. 6, 280–293.
- Selman, C., Phillips, T., Staib, J.L., Duncan, J.S., Leeuwenburgh, C., Speakman, J.R., 2005. Energy expenditure of calorically restricted rats is higher than predicted from their altered body composition. Mech. Ageing Dev. 126, 783–793.
- Selman, C., McLaren, J.S., Meyer, C., Duncan, J.S., Redman, P., Collins, A.R., Duthie, G.G., Speakman, J.R., 2006. Life-long vitamin C supplementation in combination with cold exposure does not affect oxidative damage or lifespan in mice, but decreases expression of antioxidant protection genes. Mech. Ageing Dev. 127, 897–904
- Selman, C., Lingard, S., Choudhury, A.I., Batterham, R.L., Claret, M., Clements, M., Ramadani, F., Okkenhaug, K., Schuster, E., Blanc, E., Piper, M.D., Al-Qassab, H., Speakman, J.R., Carmignac, D., Robinson, I.C., Thornton, J.M., Gems, D., Partridge, L., Withers, D.J., 2008a. Evidence for lifespan extension and delayed age-related biomarkers in insulin receptor substrate 1 null mice. FASEB J. 22, 807–818.
- Selman, C., Lingard, S., Gems, D., Partridge, L., Withers, D.J., 2008b. Comment on "Brain IRS2 signaling coordinates life span and nutrient homeostasis". Science 320, 1012.
- Semsei, I., Rao, G., Richardson, A., 1989. Changes in the expression of superoxide dismutase and catalase as a function of age and dietary restriction. Biochem. Biophys. Res. Commun. 164, 620–625.
- Shibamura, A., Ikeda, T., Nishikawa, Y., 2009. A method for oral administration of hydrophilic substances to *Caenorhabditis elegans*: effects of oral supplementation with antioxidants on the nematode lifespan. Mech. Ageing Dev. 130, 652–655.
- Sinclair, D.A., 2005. Toward a unified theory of caloric restriction and longevity regulation. Mech. Ageing Dev. 126, 987–1002.
- Smith Jr., D.L., McClure, J.M., Matecic, M., Smith, J.S., 2007. Calorie restriction extends the chronological lifespan of Saccharomyces cerevisiae independently of the Sirtuins. Aging Cell 6, 649–662.
- Song, Y., Cook, N.R., Albert, C.M., Van Denburgh, M., Manson, J.E., 2009. Effects of vitamins C and E and beta-carotene on the risk of type 2 diabetes in women at

- high risk of cardiovascular disease: a randomized controlled trial. Am. J. Clin. Nutr. 90, 429–437.
- Southam, C.M., Ehrlich, J., 1943. Effects of extract of western red-cedar heartwood on certain wood-decaying fungi in culture. Phytopathology 33, 517–524.
- Speakman, J.R., Selman, C., McLaren, J.S., Harper, E.J., 2002. Living fast, dying when? The link between aging and energetics. J. Nutr. 132, 1583S-1597S.
- Sreekumar, R., Unnikrishnan, J., Fu, A., Nygren, J., Short, K.R., Schimke, J., Barazzoni, R., Nair, K.S., 2002. Effects of caloric restriction on mitochondrial function and gene transcripts in rat muscle. Am. J. Physiol. Endocrinol. Metab. 283, E38–E43.
- Steger, R.W., Bartke, A., Cecim, M., 1993. Premature ageing in transgenic mice expressing different growth hormone genes. J. Reprod. Fertil. Suppl. 46, 61– 75
- Taguchi, A., Wartschow, L.M., White, M.F., 2007. Brain IRS2 signaling coordinates life span and nutrient homeostasis. Science 317, 369–372.
- Tapia, P.C., 2006. Sublethal mitochondrial stress with an attendant stoichiometric augmentation of reactive oxygen species may precipitate many of the beneficial alterations in cellular physiology produced by caloric restriction, intermittent fasting, exercise and dietary phytonutrients: 'Mitohormesis' for health and vitality. Med. Hypotheses 66, 832–843.
- Tatar, M., Kopelman, A., Epstein, D., Tu, M.P., Yin, C.M., Garofalo, R.S., 2001. A mutant Drosophila insulin receptor homolog that extends life-span and impairs neuroendocrine function. Science 292, 107–110.
- Tatsuta, T., Langer, T., 2008. Quality control of mitochondria: protection against neurodegeneration and ageing. EMBO J. 27, 306–314.
- Thierbach, R., Schulz, T.J., Isken, F., Voigt, A., Mietzner, B., Drewes, G., von Kleist-Retzow, J.C., Wiesner, R.J., Magnuson, M.A., Puccio, H., Pfeiffer, A.F., Steinberg, P., Ristow, M., 2005. Targeted disruption of hepatic frataxin expression causes impaired mitochondrial function, decreased life span, and tumor growth in mice. Hum. Mol. Genet. 14, 3857–3864.
- Trifunovic, A., Larsson, N.G., 2008. Mitochondrial dysfunction as a cause of ageing. J. Intern. Med. 263, 167–178.
- van Heemst, D., Beekman, M., Mooijaart, S.P., Heijmans, B.T., Brandt, B.W., Zwaan, B.J., Slagboom, P.E., Westendorp, R.G., 2005. Reduced insulin/IGF-1 signalling and human longevity. Aging Cell 4, 79–85.
- Van Raamsdonk, J.M., Hekimi, S., 2009. Deletion of the mitochondrial superoxide dismutase sod-2 extends lifespan in *Caenorhabditis elegans*. PLoS Genet. 5, e1000361.
- Vanfleteren, J.R., De Vreese, A., 1995. The gerontogenes age-1 and daf-2 determine metabolic rate potential in aging *Caenorhabditis elegans*. FASEB J. 9, 1355–1361.
- Veal, E.A., Day, A.M., Morgan, B.A., 2007. Hydrogen peroxide sensing and signaling. Mol. Cell. 26, 1–14.
- Virtamo, J., Edwards, B.K., Virtanen, M., Taylor, P.R., Malila, N., Albanes, D., Huttunen, J.K., Hartman, A.M., Hietanen, P., Maenpaa, H., Koss, L., Nordling, S., Heinonen, O.P., 2000. Effects of supplemental alpha-tocopherol and betacarotene on urinary tract cancer: incidence and mortality in a controlled trial (Finland). Cancer Causes Control 11, 933–939.
- Vivekananthan, D.P., Penn, M.S., Sapp, S.K., Hsu, A., Topol, E.J., 2003. Use of antioxidant vitamins for the prevention of cardiovascular disease: metaanalysis of randomised trials. Lancet 361, 2017–2023.
- Warburton, D.E., Nicol, C.W., Bredin, S.S., 2006. Health benefits of physical activity: the evidence. Can. Med. Ass. J. (CMAJ) 174, 801–809.
- Ward, N.C., Wu, J.H., Clarke, M.W., Puddey, I.B., Burke, V., Croft, K.D., Hodgson, J.M., 2007. The effect of vitamin E on blood pressure in individuals with type 2 diabetes: a randomized, double-blind, placebo-controlled trial. J. Hypertens 25, 227–234.
- Weindruch, R., 2006. Will dietary restriction work in primates? Biogerontology 7, 169–171
- Weindruch, R., Walford, R.L., 1988. The Retardation of Aging and Disease by Dietary Restriction. Charles C Thomas Pub Ltd., Springfield, Illinois.
- Westbrook, R., Bonkowski, M.S., Strader, A.D., Bartke, A., 2009. Alterations in oxygen consumption, respiratory quotient, and heat production in long-lived GHRKO and Ames dwarf mice, and short-lived bGH transgenic mice. J. Gerontol. A Biol. Sci. Med. Sci. 64, 443–451.
- Wick, A.N., Drury, D.R., Nakada, H.I., Wolfe, J.B., 1957. Localization of the primary metabolic block produced by 2-deoxyglucose. J. Biol. Chem. 224, 963–969.
- Wiederkehr, A., Wollheim, C.B., 2006. Minireview: implication of mitochondria in insulin secretion and action. Endocrinology 147, 2643–2649.
- Wolff, S., Dillin, A., 2006. The trifecta of aging in *Caenorhabditis elegans*. Exp. Gerontol.
- Wullschleger, S., Loewith, R., Hall, M.N., 2006. TOR signaling in growth and metabolism. Cell 124, 471–484.
- Xia, E., Rao, G., Van Remmen, H., Heydari, A.R., Richardson, A., 1995. Activities of antioxidant enzymes in various tissues of male Fischer 344 rats are altered by food restriction. J. Nutr. 125, 195–201.
- Yechoor, V.K., Patti, M.E., Ueki, K., Laustsen, P.G., Saccone, R., Rauniyar, R., Kahn, C.R., 2004. Distinct pathways of insulin-regulated versus diabetes-regulated gene expression: an in vivo analysis in MIRKO mice. Proc. Natl. Acad. Sci. USA 101, 16525–16530.
- Yen, K., Mobbs, C.V., in press. Evidence for only two independent pathways for decreasing senescence in *Caenorhabditis elegans*. Age (Dordr). doi: 10.1007/ s11357-009-9110-7.
- Yen, K., Patel, H.B., Lublin, A.L., Mobbs, C.V., 2009. SOD isoforms play no role in lifespan in ad lib or dietary restricted conditions, but mutational inactivation of SOD-1 reduces life extension by cold. Mech. Ageing Dev. 130, 173–178.

- Youngman, L.D., Park, J.Y., Ames, B.N., 1992. Protein oxidation associated with aging is reduced by dietary restriction of protein or calories. Proc. Natl. Acad. Sci. USA 89, 9112–9116.
- Zarse, K., Schulz, T.J., Birringer, M., Ristow, M., 2007. Impaired respiration is positively correlated with decreased life span in *Caenorhabditis elegans* models of Friedreich Ataxia. FASEB J. 21, 1271–1275.
- Zhang, Y., Ikeno, Y., Qi, W., Chaudhuri, A., Li, Y., Bokov, A., Thorpe, S.R., Baynes, J.W., Epstein, C., Richardson, A., Van Remmen, H., 2009. Mice deficient in both Mn superoxide dismutase and glutathione peroxidase-1 have increased oxidative damage and a greater incidence of pathology but no reduction in longevity. J. Gerontol. A Biol. Sci. Med. Sci. 64, 1212–1220.
- Zhu, Z., Jiang, W., McGinley, J.N., Thompson, H.J., 2005. 2-Deoxyglucose as an energy restriction mimetic agent: effects on mammary carcinogenesis and on mammary tumor cell growth in vitro. Cancer Res. 65, 7023–7030.
- Zid, B.M., Rogers, A.N., Katewa, S.D., Vargas, M.A., Kolipinski, M.C., Lu, T.A., Benzer, S., Kapahi, P., 2009. 4E-BP extends lifespan upon dietary restriction by enhancing mitochondrial activity in Drosophila. Cell 139, 149–160.
- Zimmerman, J.A., Malloy, V., Krajcik, R., Orentreich, N., 2003. Nutritional control of aging. Exp. Gerontol. 38, 47–52.
- Zou, S., Sinclair, J., Wilson, M.A., Carey, J.R., Liedo, P., Oropeza, A., Kalra, A., de Cabo, R., Ingram, D.K., Longo, D.L., Wolkow, C.A., 2007. Comparative approaches to facilitate the discovery of prolongevity interventions: effects of tocopherols on lifespan of three invertebrate species. Mech. Ageing Dev. 128, 222–226.
- Zureik, M., Galan, P., Bertrais, S., Mennen, L., Czernichow, S., Blacher, J., Ducimetiere, P., Hercberg, S., 2004. Effects of long-term daily low-dose supplementation with antioxidant vitamins and minerals on structure and function of large arteries. Arterioscler Thromb Vasc. Biol. 24, 1485–1491.